Taming the Fire of Nephrotoxic Botanicals

Francesca Holden^{a,b}, Vanisha Amin^{a,b}, Dominic Kuek^{a,b}, Jeffrey B. Kopp^c, Bruce M. Hendry^a, Qi-He Xu^a

^aDepartment of Renal Medicine, Centre for Integrative Chinese Medicine, School of Immunology and Microbial Sciences, Faculty of Life Sciences and Medicine, King's College London, Western Education Centre, ^bGKT School of Medical Education, Faculty of Life Sciences and Medicine, King's College London, London, United Kingdom, ^cKidney Disease Section, NIDDK, National Institutes of Health, Bethesda, MD, USA

Abstract

Criteria for diagnosing nephropathy and urothelial neoplasms induced by botanicals containing aristolochic acids (AAs) are well established. Highlights of recent research on AAs include mechanisms of AA intrarenal transport and metabolism and vigorous debate on whether AAs may also cause liver cancers. Many other botanicals may also cause renal injury, but a generalized framework for diagnosing botanical-induced kidney injury (BIKI) is lacking. Based on what we have learnt about the wide spectrum of phenotypes of BIKI attributed to AAs and a recently published standardized phenotypic framework of drug-induced kidney disease, we propose that BIKI may be categorized into six phenotypes (acute kidney injury, tubular dysfunction, glomerular disorders, nephrolithiasis, chronic kidney disease, and neoplasms) and four mechanistic types (A, predictable; B, idiosyncratic; C, chronic; and D, delayed). We call for international cooperation assembling a task force to develop, refine, and regularly appraise an online BIKI database, documenting botanical use, phenotypes, mechanisms, and levels of evidence. Once established, such a database may be linked with electronic patient records and pharmacovigilance channels to generate alerts, guide clinical decision-making, direct future research, and support evidence-based regulation of herbal medicines and education of healthcare professionals and the public. Finally, to prevent BIKI, we propose that a proactive approach integrating the triad of botanicals, users, and stakeholders will be needed.

Keywords: Adverse effects, database, herbal, kidney, mechanisms, phenotypes, prevention

INTRODUCTION

Botanicals are broadly defined as materials derived from plants and the term particularly refers to medicinal plants (herbs), herbal materials, herbal preparations, and finished herbal products that contain parts of plants as active ingredients and are used as herbal medicines or functional food. In past decades, globalization has accelerated exchanges of herbal traditions, and herbal medicines have gained popularity worldwide and become more important to the medical and pharmaceutical community, as well as to the public.^[1,2] Also known as botanical medicine, phytomedicine, or phytotherapy, herbal medicines have deep cultural roots and long historic records of use in humans. These materials are arguably the most important part of traditional medicine.^[3,4] Indeed, herbal medicines have some drug-like properties, are a rich resource and knowledge base for new drugs, and offer new options for unmet medical needs.^[5,6] In keeping with this, regulators worldwide have increasingly approved and regulated herbal medicines as drugs.[7,8]



However, herbal medicines, and other traditional medicines, can be both friends and foes to the kidneys, as elegantly reviewed by Wojcikowski *et al.*^[9,10] and most recently by Stanifer *et al.*^[11] The US National Kidney Foundation has suggested renal patients to "avoid all herbal supplements," especially 17 potential nephrotoxic herbs and over 50 phosphate-or potassium-rich herbs.^[12] While acknowledging the potential for harm, we feel that there is potential clinical and scientific value for herbal medicines in prevention and treatment of acute kidney injury (AKI)^[13] and chronic kidney disease (CKD),^[14,15]

Address for correspondence: Dr. Qihe Xu, Department of Renal Medicine, Centre for Integrative Chinese Medicine, School of Immunology and Microbial Sciences, Faculty of Life Sciences and Medicine, King's College London, Western Education Centre, Cutcombe Road, London SE5 9RJ, United Kingdom. E-mail: qihe.xu@kcl.ac.uk

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

© 2019 World Journal of Traditional Chinese Medicine | Published by Wolters Kluwer - Medknow

Received: 24-08-2018, Accepted: 04-01-2019

How to cite this article: Holden F, Amin V, Kuek D, Kopp JB, Hendry BM, Xu QH. Taming the fire of nephrotoxic botanicals. World J Tradit Chin Med 2019;5:151-63.

including drug-induced nephrotoxicity and that further research is warranted.^[16] For example, in a large cohort study of 24,971 CKD patients, use of Chinese herbal medicines was associated with a significantly better prognosis,^[17] and GQ5, a Smad3 inhibitor of botanical origin, inhibited renal fibrosis in animal models.^[18] On the other hand, as major organs for excretion of drugs and toxins, kidneys feature extensive vasculature, rich blood flow, large endothelial surface area, active tubular transport and metabolism, drastic changes of osmosis, pH, and oxygen tension^[19-21] and are particularly susceptible to injury induced by drugs and toxins, including herbal toxicants. Thus, caution must be exercised to avoid unnecessary exposure of patients, especially those with kidney diseases, to herbal medicines and other botanical materials.

Guided by the consensuses of the GP-TCM consortium 2009-2012,^[22-25] an EU-funded European-Chinese collaboration dedicated to good practice in research of traditional Chinese medicine (TCM), the corresponding author of this paper has offered medical students at King's College London three Student-Selected-Component (SSC) modules on the opportunities and challenges botanicals bring about. One of such SSC modules is designed for Year-3 graduate students to review on nephrotoxic botanicals. The SSC reviews clearly pointed to aristolochic acid nephropathy (AAN) and AA-induced neoplasms affecting the kidneys and the other parts of the urinary tract as the most established form of botanical-induced kidney injury (BIKI). This has been elegantly reviewed by Debelle et al., highlighting AA-containing species, herbs adulterated or contaminated by AA-containing species, and the worldwide spread of this problem.^[26] Yang *et al.* reviewed the variable clinical presentations of AAN,^[27] and Gökmen et al. summarized the epidemiology, diagnosis, and clinical management.^[28] More recently, AAN and AA-induced urothelial cancer and related mechanisms were updated by Jadot et al.[29] and Baudoux and Nortier.^[19] Readers are recommended to these excellent papers for details.

BIKI, however, goes beyond AAN and AAs-induced neoplasms, as reviewed by a number of experts and summarized in Table 1. Most of these peer-reviewed papers have tables listing a range of botanicals which might be "associated" with nephrotoxicity, but they often suffer from obscure evidence for causal relations and are complicated with varying phenotypic and mechanistic frameworks. Building on databases, guidelines, and publications of the GP-TCM project and the aforementioned reviews, we have conducted further reviews to inform the best strategies for preventing BIKI. We searched PubMed using the following searching strategies: ("herbal medicine" [MeSH Terms] OR traditional medicine OR phytomedicine OR phytotherapy OR Botanical* OR aristolochi*) AND (kidney* OR renal) AND (adverse effects OR side effects OR toxicity OR nephrotoxic*), and this was supplemented by more targeted literature search on specific issues. For example, "aristolochic" was searched in PubMed, and all titles published in the recent 5 years were gone through to identify noteworthy developments on AAN and AAs-induced neoplasms. Moving forward, we propose three keys: (i) Learning from the lessons of AAN and AAs-induced neoplasms; (ii) Building a reliable database; and (iii) Developing an integrative approach to prevention.

Learning from the Lessons of Aristolochic Acid Nephropathy and Aristolochic Acid-Induced Neoplasms

AAN and AA-induced neoplasms are not only the most well-established BIKI but also an excellent example to illustrate the complexity of BIKI. It can be caused by ingestion of food or drugs, knowing or unknowingly, with a range of phenotypes, including AKI, CKD, tubular disorders, and neoplasms. AAs and analogs are mainly contained in the Aristolochia and Asarum genera. According to a search of Medicinal Plant Names Service searching portal, the genus Aristolochia alone has >400 species, of which 103 have documented medicinal use worldwide and the Asarum genus has 155 species and 33 have documented medicinal use historically. In fact, the first use of an Aristolochia species - to stimulate the expulsion of the placenta during childbirth - was responsible for coining the name "Aristos lokos" or "excellent delivery," while Asarum plants were also widely used in Europe for the treatment of gout and arthralgia.[19]

Not all Aristolochia and Asarum plants are equally dangerous. In population-based case-control studies from Taiwan, Lai et al. established consumption of herbal medicines containing Guan Mu Tong (roots of Aristolochia manshuriensis) and Guang Fangchi (roots of Aristolochia *fangchi*) but not Xi Xin (roots of *Asarum heterotropoides*) and Ma Dou Ling (dried fruits of Aristolochia contorta or Aristolochia debilis), as dose-dependent independent risk factors of developing end-stage renal disease (ESRD) and urothelial neoplasms.^[30,31] In these reports, evidence is the strongest for Guan Mu Tong to cause AAN and urinary tract cancers, followed by Guang Fangchi. Accumulative ingestion of >61-100, 101-200, and >200 g Guan Mu Tong (A. manshuriensis) significantly increased odds ratio (OR) of ESRD (1.47, 2.14, and 5.82), with accumulative ingestion of Guan Mu Tong >200 g on a par with other ESRD risk factors such as diabetes (OR 4.90) and hypertension (OR 6.95).

In the aforementioned study by Lai *et al.*, ingestion of 1–30 and 31–60 g Guan Mu Tong was not significantly associated with increased ESRD risk at the population level. However, such data must be explained with care at the individual level. In light of variations in AA analog concentrations in different herbal preparations and genetic factors underlying individual susceptibility to AAs, a "safe dose" of AA analogs and AA-containing botanical products cannot yet be established despite the apparent dose dependency of AAN. For example, Guan Mu Tong contains about 25% more AAs than Guang

Review	Summary of contents and resources provided
Isnard Bagnis C, Deray G, Baumelou A, Le Quintrec M, Vanherweghem JL. Herbs and the kidney. Am J Kidney Dis 2004;44:1-11.	An overview of botanical nephrotoxicity and BIKI A summary to the clinical manifestations of BIKI, including tubular necrosis, acute interstitial nephritis, Fanconi syndrome, hypokalemia or hyperkalemia, hypertension, papillary necrosis, chronic interstitial nephritis, nephrolithiasis, urinary retention and cancer of the urinary tract Warning that some herbal medicine and impurities might affect blood pressure, drug and potassium blood levels and coagulation, induce rhabdomyolysis in renal patients; and A series of websites for further information about BIKI
Wojcikowski K, Johnson DW, Gobé G. Medicinal herbal extracts - Renal friend or foe? Part one: The toxicities of medicinal herbs. Nephrology (Carlton) 2004;9:313-8.	Three possible causes of BIKI: Phytotoxins such as AAs, impurities, herb-drug interaction Warning that chronic types of BIKI caused by insidious damage is a concern because chronic nephrotoxicity of many herbs has not been rigorously tested
Luyckx VA, Naicker S. Acute kidney injury associated with the use of traditional medicines. Nat Clin Pract Nephrol 2008;4:664-71.	Focused discussions on pathologies, manifestations, and mechanisms underlying AKI associated with traditional medicines, for example, ATN, AIN, hepatorenal syndrome, rhabdomyolysis, renal tubular acidosis, Fanconi syndrome, diabetes insipidus, papillary necrosis, transplant rejection, renal cysts, kidney stones and urinary tract obstruction, glomerular injury, urothelial malignancy, as well as signs of chronic lesions such as renal fibrosis Two online supplementary tables enlisting traditional medicines associated with AKI and CKD, respectively
Baudoux T, Nortier JL. Nephrotoxicity of herbal products. In: Toxicology of Herbal Products. Cham, Switzerland: Springer International Publishing; 2017. p. 307-44.	Nephrotoxicity of herbal products retrieved from English literature categorized into nine systemic, renal and urological syndromes, i.e., hypertension, ATN, AIN, tubular functiona disorders (Fanconi syndrome), papillary necrosis, chronic interstitial nephritis, urinary retention, urolithiasis, and urothelial cancer
	Five tables enlisting case reports on BIKI associated with intrinsic botanical nephrotoxicity herbal misuse, contamination of herbal products by drugs and heavy metals, herbal misidentification and adulteration, and herbal-drug interaction, respectively; and An introduction to the strengths and limitations of <i>in silico, in vitro, in vivo</i> , and omic methodologies for assessing renal toxicity
Jha V. Herbal medicines and chronic kidney disease. Nephrology (Carlton) 2010;15 Suppl 2:10-7.	A summary of 12 groups of plants associated with CKD, including those containing AAs, nordihydroguaiaretic acid, ephedrine, djenkolic acid, oxalic acid, glycyrrhizin, salicin, yohimbine, anthraquinone, arabinogalactan, and those contaminated by heavy metals
Xu XL, Yang LJ, Jiang JG. Renal toxic ingredients and their toxicology from traditional Chinese medicine. Expert Opin Drug Metab Toxicol 2016;12:149-59.	A summary to nephrotoxicants found in herbs used in TCM, including nephrotoxic AAs, alkaloids, anthraquinones, flavonoids, and glycosides
Stanifer JW, Kilonzo K, Wang D, Su G, Mao W, Zhang L, et al. Traditional medicines and kidney	A review on the medicinal use of botanicals in sub-Saharan African countries, China, India and Latin American countries
disease in low- and middle-income countries: Opportunities and challenges. Semin Nephrol 2017;37:245-59.	Examples to illustrate the similarities and differences of the herbal traditions and outstanding problems, highlighting low levels of evidence on efficacy and nephrotoxicity of herbal medicines tis, CKD: Chronic kidney disease, TCM: Traditional Chinese medicine, AAs: Aristolochic acid

Table 1: Recommended readings: Published reviews on botanical-induced kidney injury

ATN: Acute tubular necrosis, AIN: Acute interstitial nephritis, CKD: Chronic kidney disease, TCM: Traditional Chinese medicine, AAs: Aristolochic acids, BIKI: Botanical-induced kidney injury, AKI: Acute kidney injury

Fangchi, with Xi Xin containing only 0.5%–2% of the AAs of Guang Fangchi. These differences could explain why the former is more potently nephrotoxic and carcinogenic and why the latter does not have observable toxicity at the population level. Interestingly, cumulative ingestion of 1–1000 g Xi Xin was consistently associated with significantly reduced risk of ESRD (OR: 0.41-0.79). Thus, although Guan Mu Tong and Guang Fangchi have been banned for medicinal use worldwide, Ma Dou Ling and its corresponding aerial parts known as Tian Xian Teng, which has 10-times higher AA-I content than that of Xi Xin, remain legal herbal drugs in China, carrying warnings on its AA content and potential nephrotoxicity. For Xi Xin, despite the requirement for legal AA-I levels, it does not carry a nephrotoxic warning in the Chinese Pharmacopoeia 2015. However, water decoctions are 10-40-fold less efficient than methanol to extract AA-I from Xi Xin. Furthermore, the roots of the plant – the legal portion of the plant for medicinal use – contain 2–4-fold less AA-I amount than its aerial portions. Thus, use of the wrong plant portions including whole plant and altered extraction methods (e.g., alcoholic extract) could still lead to poisoning.^[32]

AAs have many naturally occurring analogs. It has been believed that AA I is the most toxic, followed by AA II, AA VIIIa, and AA Ia.^[33] A metabolomic analysis was carried out on 43 medicinally used *Aristolochia* species. Compounds AA I and AA II were found to be the most common AA analogs found in these extracts. AA IV, aristolactam I, and aristolactam BI were also widespread. The cytotoxicity and genotoxicity of 28 *Aristolochia* extracts were measured in HK-2 human kidney proximal tubular cells. Contrary to the prediction, no correlation was found between the amounts of AA I or AA II and extract cytotoxicity against HK-2 cells. The genotoxicity and cytotoxicity of the extracts could be linked to their contents of aristolactam BI, AA D, and AA IIIa. These results undermine the assumption that AA I and AA II are exclusively, or even chiefly, responsible for the toxicity of AA-containing species. Other analogs need to be considered as nephrotoxic agents.^[34] In this context, it should be noted that AA metabolites such as aristolactam I nitrenium ion, which readily forms AA-DNA adducts, are associated with increased tumor risk.^[29]

Intriguingly, individuals with comparable AA exposure differ greatly in developing AAN. For example, of the Belgian patients treated with the same AA-containing slimming regimen, only 2%-10% experienced AAN and those affected had varying rates of disease progression.^[35,36] Similarly, in the area where Balkan nephropathy is endemic, only 3%–7% of the population chronically exposed to AAs in food contaminated by seeds of *Aristolochia clematitis* developed AAN.^[37]

Interindividual differences in intrarenal AA transport, AA-metabolizing enzymes, and/or defense mechanisms against AAN may, at least in part, explain individual differences in resistance and susceptibility to AAN and AA-induced neoplasms. These differences could be due to genetic variations in individuals or various environmental factors that modulate enzymatic activity. Identifying these factors could potentially help in identifying vulnerable patient groups and guide treatment. For example, proximal tubular epithelial cells are the most sensitive to AAs. In these cells, organic anion transporter 1 (OAT1) plays a major role in absorbing AAs, and in HEK293 cells, an OAT1 inhibitor significantly reduced the level of AA I accumulation, while AAs induce increased apoptosis in OAT1-transfected HEK293 cells.^[38] More than 50 single nucleotide polymorphisms have been identified in the coding region of OAT1,^[39] and it would be interesting to examine if expression levels, activities and genetic variations in the OAT1 gene, and other genes involved in intrarenal AA transport affect susceptibility to AAs.

Further, cytochromes P450 (CYP) A1 and A2 enzymes are responsible for both reductive activation and oxidative detoxification of AAs. AA I is reduced at low oxygen concentrations by CYP1A1 and 1A2 and oxidized under aerobic conditions.^[40] As such, oxygen concentration in tissues may account for the differences in cytotoxicity of AAs. CYP genes are differentially expressed in males and females, at least in part due to sexual hormones, and also exhibit genetic polymorphisms. Furthermore, the activity of CYP1A1 and CYP1A2 can also be induced or inhibited by various compounds that naturally occur in fruits and vegetables.^[41] For example, baicalin, a flavone glycoside, has been shown to induce Cyp1a1 and Cyp1a2 expression in mice and attenuate AA-induced renal injury.^[42] Therefore, genetic and environmental factors that can modulate CYP1A1 and CYP1A2 activities must be further studied to understand individual susceptibility and resistance to AAN and carcinogenesis. Other enzymes that play a role on AA metabolism and susceptibility to AAs-induced BIKI include glutathione S-transferase theta 1,^[43,44] NAD(P)H quinone oxidoreductase,^[45-49] sulfotransferase 1A1,^[50,51] and cyclooxygenase/prostaglandin H synthase.^[52]

AAs and its analogs also have the potential to damage various tissues.^[53] In zebrafish embryos, AAs induce inflammation-mediated heart failure.^[54] In dogs, AA I can induce premalignant alterations in liver.^[53] In rats, AAs exhibit significant toxicity to both liver and kidneys^[55] and induce mutation of the H-Ras proto-oncogene in stomach.[56] Based on a signature A: T > T: A nucleotide substitutions, AAs and their derivatives were recently implicated in liver cancers in Taiwan and throughout Asia,^[57] but this study was widely criticized due to uncontrolled confounding factors and nonspecificity of A: T>T: A nucleotide substitution as a marker of AA exposure.^[58] In a later report from mainland China, similar mutation was not commonly observed in patients with hepatic carcinoma,[59] and indeed, genes regulate proliferation and carcinogenesis in a highly tissue-specific manner.^[60] Nonetheless, it is clear that AAs induce nephrotoxicity and cancers of the urinary tract, and exposure to AAs from all sources should be avoided.

BUILDING A RELIABLE DATABASE

Further complexity of BIKI lies in the fact that AAs and their analogs are far from the only botanical compounds that cause BIKI, and the phenotypic and mechanistic spectra of BIKI go far beyond AAN and AAs-induced neoplasms. With regard to the reviews summarized in Table 1, each contains excellent expert opinions on one aspect of BIKI, but they all more or less are limited in the following ways: (i) nonstandardized phenotypes; (ii) often unknown mechanisms; and (iii) often obscure confidence levels of causality. To address these problems, at the 16th Consortium for Globalization of Chinese Medicine Meeting held in Guangzhou, China, in August 2017, we proposed that an integrated database, linking botanicals and usage, phenotypes, mechanisms, as well as evidence and confounding factors, is desperately needed [Figure 1]. If such a database is established, the following will be possible: electronic health and medical history can be linked with nephrotoxicity databases to generate alerts to subscribers, as recently proposed by Goldstein;^[61] a consensus framework for phenotypes, mechanisms, and evidence will help with evidence-based regulation;^[62] and equally important, it will guide future research for better evidence. Herein, we interpret our proposal as follows.

Botanicals and usage

As we have learnt from AAN, whether BIKI occurs may depend on many variables: (1) whether particular species and plant parts are used, (2) whether they are processed and manufactured to meet quality standards, (3) whether their quality is compromised by contamination, adulteration, or expiry, (4) whether they are prescribed by properly trained practitioners to the right person based on right diagnoses, (5) whether the dosage and duration are appropriate, and (6) whether adverse herb-herb interaction and herb-drug interaction exist, etc. In TCM, for example, herbal medicines

Botanicals and usage Acute kidney **Urolithiasis** injury/disease (AKI/AKD) Direct cytotoxicity (A) Immune and inflammatory **Mechanisms** responses (A) Tubular of BIKI Idiosyncratic (B) dysfunction PHENOTYPES Fibrogenesis (C) (Types A-C; of BIKI Carcinogenesis (D) offending Glomerular Blockading renal transport, chemicals) disorder inducing hyperoxaluria, or causing hypertension (A/C)Chronic kidney Neoplasm disease (CKD) **Evidence & confounding factors**

Figure 1: An integrated database of BIKI and its expected main elements. Phenotypes in red fonts are those proposed by Mehta *et al.* for drug-induced kidney disease; those in black fonts are additional ones that we propose to add

should be prescribed based on TCM diagnosis and adjusted as the patients' clinical manifestations change, but they are often taken without proper guidance by qualified TCM practitioners. Thus, proper documentation of relevant information about botanicals and usage (including traditional use) is the first step of building a useful database on BIKI.

Phenotypes

Except for the particular cases of AAN and AAs-induced upper urinary tract urothelial cancer (UUC), no BIKI diagnostic framework is universally accepted. In the past decade, the International Serious Adverse Events Consortium has become a leading force in standardizing diagnosis of drug-induced adverse effects and understanding the underlying genetic mechanisms. As a not-for-profit biomedical research organization, the consortium comprises academic institutions, pharmaceutical companies, and biomedical charities and receives scientific and strategic input from the US Food and Drug Administration (FDA) and other international regulatory bodies. A panel of nephrologists and pharmacists from five different countries recently defined four standardized phenotypes of drug-induced kidney disease (DIKD), i.e., AKI, tubular dysfunction, glomerular disorders, and nephrolithiasis, along with primary and secondary clinical criteria to support the phenotype definition and a time course based on the Kidney Disease Improving Global Outcome (KDIGO)/AKI Network definitions of AKI and CKD.^[63] These phenotypes have provided a consistent framework to evaluate drug nephrotoxicity across various settings and could be borrowed for diagnosis of BIKI, but they failed to include some important phenotypes of BIKI, such as CKD and UUC, which are well-established phenotypes of AA-induced BIKI.^[27] Thus, we propose that we modify the framework into six phenotypes, by adding CKD and neoplasms. The six phenotypes of BIKI will allow mixed phenotypes, stratification, and addition of new phenotypes in the future.

Acute kidney injury

This phenotype is based on the KDIGO definition,^[64] with minor modification.^[63] AKI encompasses a rise in serum creatinine levels and mainly includes botanical-induced acute tubular necrosis (ATN) and acute interstitial nephritis (AIN) although prerenal, postrenal, and other intrarenal causes are

also possible.^[63] Although the term acute renal failure has largely been replaced by AKI,^[65] the former term is still used in reference to very serious AKI for which renal replacement therapy is needed. Indeed, the Risk, Injury, and Failure; and Loss of kidney function: and End-stage kidney disease (RIFLE) criteria and its pediatric version RIFLE recommend to grade AKI into different stages, and diagnosis at the "Risk" stage can be made before "Injury," "Failure," "Loss," and "End-stage" of renal failure are established. For specificity purposes, the phenotypic criteria proposed by Mehta et al. are stricter.^[63] For screening, alerting, and preventing purposes, however, whether AKI induced by botanicals should be proposed at earlier stages of AKI deserves further investigation. In view of the evolving definition of AKI and the difficulty in establishing causality, the true prevalence of BIKI manifesting as AKI remains largely unknown. In the developing countries, however, it was reported that folk remedies accounted for up to 35% of cases of AKI.^[66] We list some best-known examples of botanicals associated with AKI in Supplementary Table 1. These include AA-containing species, nephrotoxic flavonoid-containing species, triptolide-containing species, Aloe vera, Aloe ferox, Callilepis laureola (Impila), Teucrium polium, Artemisia absinthium (wormwood), and Uncaria tomentosa or U. guianensis (cat's claw), etc.

Nephrolithiasis

Renal calculi develop most commonly from calcium oxalate.^[67,68] Calculi provoke symptoms and signs such as renal colic, nausea, vomiting, hematuria, pyuria, pyrexia, and dysuria and may cause ureteric obstruction and reduced renal function. Some botanical compounds can precipitate as crystals, depending on their urinary solubility. Patients may be asymptomatic, or this may lead to isolated crystalluria or stones. To classify the nephrolithiasis phenotype, imaging such as ultrasound should be performed to definitively visualize a stone, with or without obstruction. In addition, due to the naturally high incidence of calculi in the population, when conducting investigations, it is imperative to establish the temporal relationship of the suspect botanical and analyze calculi composition. Furthermore, there should be no prior history of calculi as this may influence the clinical interpretations made regarding the particular botanical.^[63] This phenotype may induce AIN and/or renal tubular acidosis syndromes, and obstructive calculi could induce AKI or CKD.^[63] Of note, partial obstruction and unilateral obstruction could damage renal function without leading to a rise in serum creatinine due to the strong renal function reserve. If there is nonobstructive nephrolithiasis, then ultrasound should detect the presence of stones, or urinalysis should detect crystals unless the drug has been discontinued for some time.^[63] Many factors can influence the formation of kidney stones but in particular, hyperoxaluria and hypercalciuria are known to play a significant role.^[63] Botanicals associated with calcium oxalate calculi and nephrolithiasis include cranberry (Vaccinium species) juice, rhubarb (Rheum officinale), star fruit (Averrhoa carambola), Ephedrine (a compound isolated from Ephedra *sinica),* Guaifenesin (a constituent of *Guaiacum officinale*), etc., [Supplementary Table 2].

Tubular dysfunction

Botanicals may lead to tubular dysfunction ranging from isolated dysfunction such as phosphate wasting to more generalized damage causing acquired Fanconi syndrome, diabetes insipidus, or proximal tubular acidosis. Mehta et al. included the tubular dysfunction phenotype in their framework of DIKD. It is characterized by abnormal urinary losses of glucose, phosphate, potassium, magnesium, water, and tubular proteins and secondary abnormalities such as changes in serum electrolytes, pH, and bicarbonate. The latter must be also present to improve specificity.^[63] AA-containing botanicals are known to cause tubular dysfunction, which can manifest as Fanconi syndrome.^[69-72] Fanconi syndrome involves dysfunction of the proximal renal tubule, leading to urinary loss of glucose, amino acids, phosphate, uric acid, and bicarbonate. Other botanicals associated with tubular dysfunction include Cleistanthus collinus and diuretic Juniper berries (Juniperus communis), dandelion (Taraxacum officinale), asparagus root (Asparagus officinalis), lovage root (Levisticum officinale), parsley (Petroselinum crispum), stinging nettle leaf (Urtica dioica), etc., [Supplementary Table 3].

Glomerular disorders

Botanicals may induce glomerular injury, with patients presenting with hematuria, proteinuria, and associated urinary sediment abnormalities. Based on the DIKD framework of Mehta et al., primary criteria should include substantial proteinuria and a kidney biopsy demonstrating glomerular disease that can be plausibly associated with a particular botanical and not with another disease.^[63] Yellow oleander (Cerbera thevetia) was reported to cause hematuria and proteinuria, suggestive of glomerular injury.^[66] Animal studies demonstrated that feeding vellow oleander seeds to rats induced glomerular endothelial proliferation and glomerular hypercellularity. Further, both human and animal kidneys have displayed ATN postmortem,^[73-76] indicating tubular toxicity. In addition, arsenic-contaminated bladderwrack (Fucus vesiculosus) was reported to induce mesangial proliferation, interstitial fibrosis, and tubular degeneration, manifesting proteinuria and hematuria.[77]

Chronic kidney disease

Botanicals can induce kidney injury over a long period of time. For example, *Glycyrrhiza glabra* and ephedrine-containing *Ephedra* spp. or herbal mixtures of ephedrine-containing components may induce hypertension, a risk factor for CKD progression.^[78,79] However, AAN is probably the best example demonstrating that BIKI can manifest either acutely or chronically.^[27] An observational study which involved 300 individuals showed that AAN has variant phenotypes of BIKI, including AKI, tubular dysfunction, and CKD, with the latter being the most common clinical manifestation. The clinical subtypes were associated with cumulative doses and time course of AA consumption, for example, those exposed to the lowest dose but for the longest period of exposure to AAs.^[27] One such phenotype of AAN is found in Balkan endemic nephropathy, a chronic tubulointerstitial kidney disease found in farming villages in Bulgaria, Romania, and Serbia.^[80] It is characterized by an asymptomatic onset with a slow progression to ESRD and increased frequency of UUC. It was first suggested that environmental exposure to AA may be a cause of Balkan endemic nephropathy when Ivic found wheat flour was contaminated with A. clematitis seeds.^[81] Subsequently, aristolactam-DNA adducts and hallmark A: T > T: A transversions have been found in renal cortical and urothelial malignant tissue of patients with Balkan endemic nephropathy.[81-85] A storage protein called dioscorin in Dioscorea villosa (wild yam) increases the expression of cytokines involved in renal fibrosis, such as transforming growth factor (TGF)- beta 1.^[86-88] When administered to rats. D. villosa increases expression of renal TGF-B1 and induces renal fibrosis and hepatic inflammation after 28 days.[86] Furthermore, Leonurus japonicus (known as Yimucao in TCM) was reported to have in vitro profibrotic activities[89] and induce renal fibrosis in animal models, and its toxicity was reduced when used in TCM formulae.[90,91]

Renal and urothelial neoplasms

After the original reports of AAN, a body of evidence emerged. associating UUC with the consumption of AA-containing botanicals,[92-95] highlighting that BIKI can also manifest with renal or urothelial neoplasms. In a study, 4 out of 10 patients with AAN had a multifocal high-grade carcinoma in situ,^[96] and this has been confirmed in larger studies of patients with AAN, which show that the risk for urothelial neoplasms is associated with doses of A. fangchi.[97] A similar rate of UUC was reported in a 15-year follow-up study along with a rise in the incidence of late-onset bladder tumors.^[98,99] Studies in Taiwan have reported a very high incidence of UUC which is also associated with CKD^[100] and a marked dose-dependent relationship between the ingestion of AA-containing botanicals and the risk of UUC.^[31] AA exposure has been widely implicated in the development of UUC. Their carcinogenic effects have been well described with their ability to induce A: T > T: A transversions in the gene TP53.^[101] This transversion signature is also present at the genome-wide level.[102,103] However, an association between AA exposure and renal cell carcinomas (RCC) has not been as widely explored. A whole-genome sequencing study of RCCs found that 12 out of 14 Romanian RCC cases demonstrated high rates of A: T > T: A transversions. In contrast, this mutational signature was absent in 80 other studied cases diagnosed in Europe.^[104] In addition, the nonmalignant renal cortical tissue from the 14 Romanian cases studied did not display the hallmark histological features of AAN such as interstitial fibrosis or tubular atrophy. Therefore, these results triggered a further study to determine whether Aristolactam-DNA adducts were present in nonmalignant renal cortical tissue samples.^[105] A similar level of DNA adducts was found in the 14 Romanian cases to the renal cortical tissue of patients with UUC in Taiwan and the Balkans,^[105,106] and additionally, the proportion of A: T > T: A mutations positively correlated with the number of DNA-adducts.^[105] Other studies have found that the RCC tissue samples of 5 of 8 patients with Balkan endemic nephropathy displayed the A:T > T:A transversion mutational signature. This signature was missing in control samples of RCCs from patients of non-Balkan endemic regions.^[107]

Mechanisms

More than a quarter century ago, Huxtable enlisted a series of factors predisposing to intoxication from the use of herbs. These include the following: misidentification of a plant; the unknown or ignored toxicity of a correctly identified plants; difficulties in identifying chopped, processed herbs, or plant mixtures; persistent use of a toxic plant; variability in toxic plant constituents; problems of nomenclature; adulteration and the difficulty in establishing the chronic toxic potential of a plant; certain human populations at higher risk of intoxication, including chronic users, those consuming large amounts or a great variety, the very young, fetuses, the elderly, the sick, the malnourished and those on long-term medication, and certain ethnic groups; and certain plant toxins with gender-selective action.^[108]

Much information surrounding BIKI remains unknown. To be pragmatic, however, BIKI can be divided into the following four types based on the classification of drug adverse effects by Edwards and Aronson:^[109] (i) type A, which are attributable directly to the toxicity and properties of the botanicals themselves, thus predictable and often dose dependent; (ii) type B, which are attributable to personalized responses to botanicals, thus idiosyncratic and often not dose related or predictable by pharmacology; (iii) type C (chronic), which have cumulative effect; and type D, which have delayed onset and are carcinogenic and genotoxic. Of note, types B, C, and D are difficult to be noticed and established.

As illustrated by the case of AAN, identifying offending compounds has a critical importance in mechanistic studies of botanicals. Biologically, BIKI mechanisms can be further attributed to cytotoxicity, immune and pro-inflammatory responses, fibrogenesis, carcinogenesis, and blockade of renal transport, inducing hyperoxaluria and causing hypertension. Cytotoxicity can be due to regulated cell death mechanisms, including pyroptosis, apoptosis, and necroptosis,^[110] and un-programmed necrosis, which all play important roles in inflammation,^[111] which in turn play important roles in injury and repair. While botanical-induced apoptosis is most studied, roles for other types of cell death in BIKI remain elusive and deserve further studies. Supplementary Table 4 summarizes the current state of knowledge of BIKI biological mechanisms.

Evidence and confounding factors

Currently, many people including professionals use the internet to find medical information. Although practical, this can be misleading. Some websites^[112,113] and earlier papers^[114] labeled tens of herbs as "nephrotoxic," but many provided no source of information and supporting evidence and some could be due to adulteration or contamination. For example,

"Chaihu" (Bupleurum chinense roots) has been labeled as "nephrotoxic," but we could not find evidence in support of this claim. Indeed, "Chaihu" adulteration by nephrotoxic A. manshuriensis roots has been recently reported.^[115] Senecionis Scandentis Herba (Qianliguang, the aerial portion of Senecio scandens) has also been listed as "nephrotoxic" but evidence supporting this claim is lacking; indeed, oral administration of an aqueous extract of the herb, 225, 450, and 900 mg/kg/d, for 90 days was reported to increase serum creatinine, potassium, and chloride in some rats, without changes in blood urea nitrogen levels, gross renal histology, urinary volume, urinary glucose, bilirubin, ketone body, specific gravity, occult blood, pH, protein, urobilinogen, nitrite, and leukocyte levels.^[116] These findings suggest that the agent might affect bodily metabolism and/or impair renal tubular function. Thus, although the phytochemistry, pharmacological, and hepatotoxic properties of this pyrrolizidine alkaloid-containing herb are well documented,[117] whether it is nephrotoxic remains obscure.

Evidence of BIKI is often provided by case reports, which do not readily establish causality. Furthermore, a proportion of the evidence for BIKI mechanisms comes from animal or *in vitro* studies, which need to be carefully interpreted in terms of their clinical relevance. Thus, huge challenges often exist when assessing causal relations, in view that there are no internationally agreed standards or criteria for assessing causality in individual cases. Thus, we will restrain from labeling specific botanicals as "nephrotoxic" or any links as "causal" or "non-causal." Nonetheless, we do think it important to compare and refine existing algorithms^[118,119] and to develop new ones for calculating causality scores for BIKI, which should be based on accumulating evidence and should indicate the likelihood of any causal BIKI. Evidence from human cases is likely confounded by various factors. These include the presence of chronic conditions such as diabetes and hypertension, genetics and family history, diet, medication history, and access to healthcare. All of these may influence the mechanisms, pathways, progression, and/ or treatment of BIKI.

One confounding factor particularly worth considering is whether it is the botanical itself causing nephrotoxicity or its adulterants and contaminants, for example, heavy metals and pesticides. For example, a case report stated that Bladderwrack (Fucus vesiculosus), a brown seaweed from the Fucaceae family, induced diabetes insipidus and tubular dysfunction after 3 months daily consumption.^[77] The researchers found that the preparation was contaminated with high levels of arsenic,^[77] and therefore, its pathogenesis is likely associated with the contamination of kelp preparations with arsenic and other heavy metals, due to growth in polluted waters.^[120,121] Other case reports have described contamination of herbal products with cadmium leading to renal tubular dysfunction.^[122] However, the degree to which cadmium was completely responsible was questioned, highlighting the difficulties in elucidating whether injury is due to contaminants, the plant itself, or a combination of the two with a preparation generating a dangerous interaction. This may lead to certain botanicals being misreported as nephrotoxic in the literature.

DEVELOPING AN INTEGRATIVE AND PROACTIVE APPROACH TO PREVENTION An integrative approach

In contrast to fire, which can be stopped when any side of the oxygen-fuel-heat triangle is removed, BIKI can be

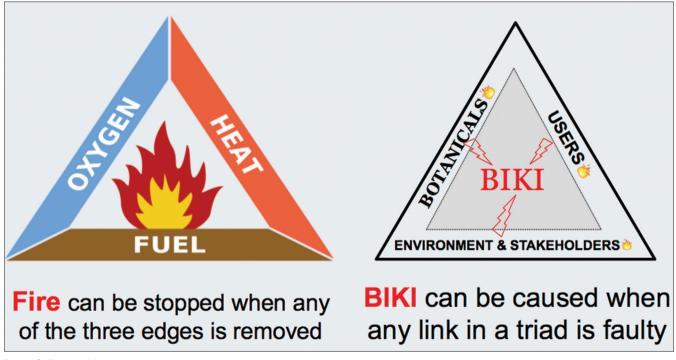


Figure 2: Botanical-induced kidney injury is trickier than fire and demands an integrated approach to prevention

triggered when any of the botanicals-users-stakeholders triads is faulty [Figure 2]. Thus, BIKI prevention demands a sophisticated approach integrating data on botanicals and usage, individualized response after exposure, as well as environment and stakeholders, including farmers, manufacturers, healthcare providers, and regulators, as proposed for herbal safety by Williamson *et al.*^[25]

First, as having been emphasized by the FP7 GP-TCM consortium^[123] and others,^[124] the nomenclature issue is critical in developing integrative evidence, an integrative database, and an integrated approach to prevention. Adverse reaction reports, whether submitted to regulatory authorities or published in the literature, are meaningless and even misleading if the medicinal herb(s) or botanical ingredients in a product cannot be identified. Names for medicinal herbs include the Latin scientific name, the common or vernacular name, the pharmaceutical name or pharmacopeial name, or the specific herbal drug names (as those used in TCM). Herbal prescriptions, product packaging, or labels may have one or more of these depending on the source and regulatory status of the product. These have to be interpreted with care as even the scientific names may have synonyms. On this matter, Kew's Medicinal Plant Names Services (MPNS)^[125] and the World Health Organization (WHO) Herbal Dictionary^[126] can be expected to play a leading role. A botanically correct label does not necessarily confirm that the product contains what is listed on the label and that the concerned botanicals are not adulterated or contaminated and are in the expected quality and quantity. In cases of serious adverse reactions where specific toxins are suspected, laboratory analysis of the product/herb may be advisable to verify the reports. Beyond naming, labeling and prescription information, processing and preparation, administration route, dosing and timing are all important factors that may affect BIKI.

Second, in an era which aims for personalized medicine, individual factors including botanical user's age, sex, pregnancy status, diet, genetic background, educational status, health status, ability and willingness to understand and adhere, self-prescription, nutritional status, and health habits can all be important factors that affect BIKI. According to TCM theories, different patients with the same diseases or the same patient at the different stages of disease may vary in terms of TCM diagnosis. Many TCM practitioners believe that botanical prescription and use guided by TCM diagnosis may minimize risk-benefit ratio and this belief deserves further investigation. Third, environment has an important place in Good Agriculture and Collection Practice and Good Manufacture Practice and profoundly affects the quality of botanical materials and products. Thus, environment, through affecting quality of botanical materials and products, plays important roles in BIKI. Fourth, the roles for stakeholders ranging from vendors, healthcare providers, regulators, scientists, professional organizations, and their interaction with botanicals and botanical users can never be overemphasized.

Proactiveness is required

A reliable database that we have proposed above will help identify new cases of suspected BIKI. Meanwhile, we must identify and avoid exposure to risks proactively. Each phenotype and identified mechanisms of BIKI may be area of focused studies for better integrative evidence, better understanding of mechanisms, better tools for diagnosis, and better prevention.

First, current literature particularly focuses on the AKI phenotype based on markers such as serum creatinine or blood nitrogen urea. However, a rise in these markers occurs only after significant kidney injury,^[127,128] suggesting the need for more sensitive and earlier detection. Serum creatinine also varies depending on age, gender, muscle mass, and nutrition,^[129] reducing its reliability. Therefore, the development of new biomarkers may lead to better understanding, diagnosis of the AKI type of BIKI, ultimately leading to better care.

For instance, heme oxygenase-1 (HO-1), which is involved in heme degradation,^[130] could be a potential biomarker for *in vitro* screening and early marker of BIKI. The function of HO-1 is not completely understood. Both in vitro and in vivo models of injury have suggested that endogenous HO-1 is cytoprotective, and chemical or genetic inhibition of HO-1 increases cell death and tissue necrosis.^[130] This is further supported by studies of HO-1 knock-out mice, whereby targeted deletion of the enzyme leads to death in utero or within 1 year of birth.^[131,132] Mice that survived beyond 1 year had many abnormalities such as growth retardation, anemia, iron deposition in the organs, and chronic inflammation such as glomerulonephritis, in addition to cells which were more susceptible to oxidative stress from endotoxins. HO-1 expression has been shown to be highly upregulated in response to cell injury mediated by oxidative or pro-inflammatory stress, heavy metals, ischemia, and hypoxia,^[130] and renal HO-1 expression is increased in animal models of many types of AKI.^[133-135] HO-1 was upregulated in the urine of patients with AKI or tubulointerstitial damage.[136] This suggested that HO-1 expression could be applied as a diagnostic tool to identify and monitor patients with kidney disease. A recent study aimed to identify a consistent biomarker of nephrotoxicity through gene expression profiling of human proximal tubular epithelial cells postexposure to different concentrations of nephrotoxicants.^[137] The gene for HO-1 was significantly induced, in a dose-dependent fashion, by 6 out of 9 nephrotoxic compounds, including AAs, highlighting its potential as a biomarker.^[137] As not all nephrotoxic compounds induce HO-1, the underlying mechanisms and other new biomarkers are surely needed.

As mentioned earlier, causal links between botanicals and CKD phenotype are difficult to establish clinically. In view that fibrosis is a cardinal feature of progressive CKD and that certain botanicals have been associated with epithelial-to-mesenchymal transition in epithelial cells and fibrogenesis in renal fibroblasts and in animal models,^[138] it is likely *in vitro*, *in vivo*, and *in silico* models for detecting

Holden, et al.

botanical-induced epithelial-to-mesenchymal transition and fibrogenesis may be useful to identify botanicals that may cause CKD, leading to required labelling or banning from the marketplace. Similarly, botanical products could be screened for other phenotypes and mechanisms of BIKI in creative models. These will collectively contribute to the success of proactive and integrative pharmacovigilance for the prevention of BIKI.

CONCLUDING REMARKS

BIKI is an important and complex societal and medical problem. Learning from past lessons, we have proposed it as priorities to develop an authoritative BIKI database and to adopt a proactive, integrated approach to prevention. Building on existing expert opinions [e.g., those summarized in Table 1], we call for international leadership and interdisciplinary cooperation toward establishing a one-stop, open-access, user-friendly, high-quality, human-curated, and regularly updated BIKI database.

Despite all the challenges, we believe that harmonization of botanical pharmacovigilance with current pharmacovigilance system designed to report adverse drug reactions (ADR) induced by pure compounds is possible. We suggest that the WHO is uniquely qualified to play a leading role in this initiative. The international stature of WHO provides the legitimacy needed for such a global issue, and the most recent election of Dr. Tedros Adhanom Ghebrevesus as the first African Director-General of this global agency may be an opportunity. Traditional medicine plays a major role in achieving universal health coverage and should be made safer and more reliable. Funding for such a long-term project could come from the member states and other fundraising channels and could include funds that currently support the Traditional, Complementary, and Integrative Medicine program of the WHO and the US National Center for Complementary and Integrative Health, for example. The initiative should involve the WHO Collaborating Centre for International Drug Monitoring, Uppsala Monitoring Centre (UMC), MPNS, national and international medical, pharmacological, toxicological, and pharmacovigilance centers and societies, as well as individuals such as experts involved in the work listed in Table 1.

Illustrative is the work of UMC, which takes ADR reports from over 100 countries around the world. The UMC database contains millions of reports, including tens of thousands related to herbal or natural products. These reports are incorporated into a single database, with review of suspected signals carried out by experts in relevant fields.^[23,139] Unfortunately, kidney-related ADR was rarely reported by UMC. This either means that kidney-related ADR is rare or more likely that nephrology practitioners are insufficiently aware of this resource.

National regulators also have an important role to play. Recently, FDA has launched a new adverse event portal that enables drug developers, doctors, and patients to search for safety red flags

for approved drugs. This FDA Adverse Event Reporting System offers a powerful postmarketing pharmacovigilance resource and a means of guiding preclinical drug development.^[140] This new portal can also serve as a potential platform for pharmacovigilance of nephrotoxic botanicals.

After all, as Theodore Roosevelt said: "Risk is like fire: if controlled it will help us; if uncontrolled it will rise up and destroy us." Botanicals are such risks. They can become valuable remedies or cause damage. It all depends on how well we know them and whether they are used appropriately.

Financial support and sponsorship

The authors would like to thank Kidney Research UK, Innovation China UK, European Union and the Intramural Research Program, NIDDK, NIH for funding of this important line of research.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Pelkonen O, Xu Q, Fan TP. Why is research on herbal medicinal products important and how can we improve its quality? J Tradit Complement Med 2014;4:1-7.
- 2. Pan SY, Gao SH, Zhou SF, Tang MK, Yu ZL, Ko KM, *et al.* New perspectives on complementary and alternative medicine: An overview and alternative therapy. Altern Ther Health Med 2012;18:20-36.
- Pan SY, Litscher G, Gao SH, Zhou SF, Yu ZL, Chen HQ, *et al.* Historical perspective of traditional indigenous medical practices: The current renaissance and conservation of herbal resources. Evid Based Complement Alternat Med 2014;2014:525340.
- 4. Xu Q, Bauer R, Hendry BM, Fan TP, Zhao Z, Duez P, *et al.* The quest for modernisation of traditional Chinese medicine. BMC Complement Altern Med 2013;13:132.
- 5. Fan T, Briggs J, Liu L, Lv A, van der Greef J, Xu A. Integrating traditional medicine into modern health care. Science 2014;346:S4.
- Xu Q, Qu F, Pelkonen O. Network Pharmacology and Traditional Chinese Medicine. In: Sakagami H, editor. Alternative Medicine. InTech. Available from: https://www.intechopen.com/books/alternativemedicine/network-pharmacology-and-traditional-chinese-medicine [Last accessed on 2019 Feb 20].
- Zhang Q, Kelly E. The WHO traditional medicine strategy 2014-2023: A perspective. Science 2014;346:S5-7.
- Fan TP, Deal G, Koo HL, Rees D, Sun H, Chen S, *et al.* Future development of global regulations of Chinese herbal products. J Ethnopharmacol 2012;140:568-86.
- Wojcikowski K, Johnson DW, Gobé G. Medicinal herbal extracts – Renal friend or foe? Part one: The toxicities of medicinal herbs. Nephrology (Carlton) 2004;9:313-8.
- Wojcikowski K, Johnson DW, Gobé G. Medicinal herbal extracts renal friend or foe? Part two: Herbal extracts with potential renal benefits. Nephrology (Carlton) 2004;9:400-5.
- Stanifer JW, Kilonzo K, Wang D, Su G, Mao W, Zhang L, *et al.* Traditional medicines and kidney disease in low – And middle-income countries: Opportunities and challenges. Semin Nephrol 2017;37:245-59.
- National Kidney Foundation Website. Herbal Supplements and Kidney Disease. Available from: https://www.kidney.org/atoz/content/ herbalsupp. [Last accessed on 2017 May 24].
- Bunel V, Qu F, Duez P, Xu Q. Herbal medicines for acute kidney injury: Evidence, gaps and frontiers. World J Tradit Chin Med 2015;1:47-66.
- Zhong Y, Deng Y, Chen Y, Chuang PY, Cijiang He J. Therapeutic use of traditional Chinese herbal medications for chronic kidney diseases. Kidney Int 2013;84:1108-18.
- 15. Zhong Y, Menon MC, Deng Y, Chen Y, He JC. Recent advances in

traditional Chinese medicine for kidney disease. Am J Kidney Dis 2015;66:513-22.

- Khajavi Rad A, Mohebbati R, Hosseinian S. Drug-induced nephrotoxicity and medicinal plants. Iran J Kidney Dis 2017;11:169-79.
- Lin MY, Chiu YW, Chang JS, Lin HL, Lee CT, Chiu GF, et al. Association of prescribed Chinese herbal medicine use with risk of end-stage renal disease in patients with chronic kidney disease. Kidney Int 2015;88:1365-73.
- Ai J, Nie J, He J, Guo Q, Li M, Lei Y, *et al.* GQ5 hinders renal fibrosis in obstructive nephropathy by selectively inhibiting TGF-β-induced smad3 phosphorylation. J Am Soc Nephrol 2015;26:1827-38.
- Baudoux T, Nortier J. Nephrotoxicity of herbal products. In: Pelkonen O, Duez P, Vuorela P, Vuorela H, editors. Toxicology of Herbal Products. Cham, Switzerland: Springer International Publishing; 2017. p. 307-44.
- Jha V. Herbal medicines and chronic kidney disease. Nephrology (Carlton) 2010;15 Suppl 2:10-7.
- Isnard Bagnis C, Deray G, Baumelou A, Le Quintrec M, Vanherweghem JL. Herbs and the kidney. Am J Kidney Dis 2004;44:1-1.
- 22. Zhang L, Yan J, Liu X, Ye Z, Yang X, Meyboom R, *et al.* Pharmacovigilance practice and risk control of traditional Chinese medicine drugs in China: Current status and future perspective. J Ethnopharmacol 2012;140:519-25.
- Shaw D, Graeme L, Pierre D, Elizabeth W, Kelvin C. Pharmacovigilance of herbal medicine. J Ethnopharmacol 2012;140:513-8.
- 24. Ouedraogo M, Baudoux T, Stévigny C, Nortier J, Colet JM, Efferth T, et al. Review of current and "omics" methods for assessing the toxicity (genotoxicity, teratogenicity and nephrotoxicity) of herbal medicines and mushrooms. J Ethnopharmacol 2012;140:492-512.
- Williamson E, Chan K, Xu Q, Nachtergael A, Bunel V, Zhang L, *et al.* Evaluating the safety of herbal medicines: Integrated toxicological approaches. Science 2015;347:S47-9.
- Debelle FD, Vanherweghem JL, Nortier JL. Aristolochic acid nephropathy: A worldwide problem. Kidney Int 2008;74:158-69.
- Yang L, Su T, Li XM, Wang X, Cai SQ, Meng LQ, *et al.* Aristolochic acid nephropathy: Variation in presentation and prognosis. Nephrol Dial Transplant 2012;27:292-8.
- Gökmen MR, Cosyns JP, Arlt VM, Stiborová M, Phillips DH, Schmeiser HH, *et al.* The epidemiology, diagnosis, and management of aristolochic acid nephropathy: A narrative review. Ann Intern Med 2013;158:469-77.
- Jadot I, Declèves AE, Nortier J, Caron N. An integrated view of aristolochic acid nephropathy: Update of the literature. Int J Mol Sci 2017;18. pii: E297.
- Lai MN, Lai JN, Chen PC, Hsieh SC, Hu FC, Wang JD, et al. Risks of kidney failure associated with consumption of herbal products containing mu tong or *Fangchi*: A population-based case-control study. Am J Kidney Dis 2010;55:507-18.
- Lai MN, Wang SM, Chen PC, Chen YY, Wang JD. Population-based case-control study of Chinese herbal products containing aristolochic acid and urinary tract cancer risk. J Natl Cancer Inst 2010;102:179-86.
- Zhao ZZ, Liang ZT, Jiang ZH, Leung KS, Chan CL, Chan HY, et al. Comparative study on the aristolochic acid I content of herba asarifor safe use. Phytomedicine 2008;15:741-8.
- Balachandran P, Wei F, Lin RC, Khan IA, Pasco DS. Structure activity relationships of aristolochic acid analogues: Toxicity in cultured renal epithelial cells. Kidney Int 2005;67:1797-805.
- Michl J, Kite GC, Wanke S, Zierau O, Vollmer G, Neinhuis C, et al. LC-MS – And (1)H NMR-based metabolomic analysis and *in vitro* toxicological assessment of 43 *Aristolochia* species. J Nat Prod 2016;79:30-7.
- Vanherweghem JL, Depierreux M, Tielemans C, Abramowicz D, Dratwa M, Jadoul M, *et al.* Rapidly progressive interstitial renal fibrosis in young women: Association with slimming regimen including Chinese herbs. Lancet 1993;341:387-91.
- 36. Stiborová M, Frei E, Wiessler M, Schmeiser HH. Human enzymes involved in the metabolic activation of carcinogenic aristolochic acids: Evidence for reductive activation by cytochromes P450 1A1 and 1A2. Chem Res Toxicol 2001;14:1128-37.
- 37. Stiborová M, Arlt VM, Schmeiser HH. Balkan endemic nephropathy:

An update on its aetiology. Arch Toxicol 2016;90:2595-615.

- Zeng Y, Zhang R, Wu J, Liu M, Peng W, Yu X, *et al.* Organic anion transporter 1 (OAT1) involved in renal cell transport of aristolochic acid I. Hum Exp Toxicol 2012;31:759-70.
- Li Z, Lam P, Zhu L, Wang K, Zhou F. Current Updates in the Genetic Polymorphisms of Human Organic Anion Transporters (OATs). J Pharmacogenomics Pharmacoproteomics 2012;3:127.
- Stiborová M, Levová K, Bárta F, Shi Z, Frei E, Schmeiser HH, et al. Bioactivation versus detoxication of the urothelial carcinogen aristolochic acid I by human cytochrome P450 1A1 and 1A2. Toxicol Sci 2012;125:345-58.
- Felicia W. In vitro Inhibition of CYP1A and CYP3A by Phenolic Compounds from Bilberry (Vaccinium myrtillus), in Male and Female Porcine Liver Microsomes. Swedish University of Agricultural Sciences;2015.
- 42. Wang K, Feng C, Li C, Yao J, Xie X, Gong L, *et al.* Baicalin protects mice from aristolochic acid I-induced kidney injury by induction of CYP1A through the aromatic hydrocarbon receptor. Int J Mol Sci 2015;16:16454-68.
- 43. Chen B, Bai Y, Sun M, Ni X, Yang Y, Yang Y, *et al.* Glutathione S-transferases T1 null genotype is associated with susceptibility to aristolochic acid nephropathy. Int Urol Nephrol 2012;44:301-7.
- 44. Ni X, Zheng S, Xu F, Sun M, Yang Y, Fu J, et al. Association of GSTT1, GSTM1 and GSTP1 gene polymorphism with aristolochic acid nephropathy. Chin J Nephrol 2005;24:614-8.
- 45. Stiborová M, Frei E, Sopko B, Sopková K, Marková V, Lanková M, *et al.* Human cytosolic enzymes involved in the metabolic activation of carcinogenic aristolochic acid: Evidence for reductive activation by human NAD(P)H: Quinone oxidoreductase. Carcinogenesis 2003;24:1695-703.
- 46. Stiborová M, Mareš J, Frei E, Arlt VM, Martínek V, Schmeiser HH, et al. The human carcinogen aristolochic acid i is activated to form DNA adducts by human NAD(P)H: Quinone oxidoreductase without the contribution of acetyltransferases or sulfotransferases. Environ Mol Mutagen 2011;52:448-59.
- 47. Chen M, Gong L, Qi X, Xing G, Luan Y, Wu Y, et al. Inhibition of renal NQO1 activity by dicoumarol suppresses nitroreduction of aristolochic acid I and attenuates its nephrotoxicity. Toxicol Sci 2011;122:288-96.
- Bárta F, Levová K, Frei E, Schmeiser HH, Arlt VM, Stiborová M, et al. The effect of aristolochic acid I on expression of NAD(P)H: Quinone oxidoreductase in mice and rats – A comparative study. Mutat Res Genet Toxicol Environ Mutagen 2014;768:1-7.
- Stiborová M, Frei E, Arlt VM, Schmeiser HH. Metabolic activation of carcinogenic aristolochic acid, a risk factor for Balkan endemic nephropathy. Mutat Res 2008;658:55-67.
- Hashimoto K, Zaitseva IN, Bonala R, Attaluri S, Ozga K, Iden CR, et al. Sulfotransferase-1A1-dependent bioactivation of aristolochic acid I and N-hydroxyaristolactam I in human cells. Carcinogenesis 2016;37:647-55.
- Meinl W, Pabel U, Osterloh-Quiroz M, Hengstler JG, Glatt H. Human sulphotransferases are involved in the activation of aristolochic acids and are expressed in renal target tissue. Int J Cancer 2006;118:1090-7.
- Stiborova M, Frei E, Breuer A, Wiessler M, Schmeiser H. Evidence for reductive activation of carcinogenic aristolochic acids by prostaglandin H synthase – 32P-postlabeling analysis of DNA adduct formation. Mutat Res Genet Toxicol Environ Mutagen 2001;493:149-60.
- 53. Jin K, Su KK, Li T, Zhu XQ, Wang Q, Ge RS, *et al.* Hepatic premalignant alterations triggered by human nephrotoxin aristolochic acid I in canines. Cancer Prev Res (Phila) 2016;9:324-34.
- Huang CC, Chen PC, Huang CW, Yu J. Aristolochic acid induces heart failure in Zebrafish embryos that is mediated by inflammation. Toxicol Sci 2007;100:486-94.
- 55. Yeh YH, Lee YT, Hsieh HS, Hwang DF. Short-term toxicity of aristolochic acid, aristolochic acid-I and aristolochic acid-II in rats. Food Chem Toxicol 2008;46:1157-63.
- Cheng CL, Chen KJ, Shih PH, Lu LY, Hung CF, Lin WC, *et al.* Chronic renal failure rats are highly sensitive to aristolochic acids, which are nephrotoxic and carcinogenic agents. Cancer Lett 2006;232:236-42.
- 57. Ng AW, Poon SL, Huang MN, Lim JQ, Boot A, Yu W, *et al.* Aristolochic acids and their derivatives are widely implicated in liver cancers in

Taiwan and throughout Asia. Sci Transl Med 2017;9. pii: eaan6446.

- Zhou G, Zhao X. Carcinogens that induce the A:T>T:A nucleotide substitutions in the genome. Front Med 2018;12:236-8.
- Ji X, Feng G, Chen G, Shi T. Lack of correlation between aristolochic acid exposure and hepatocellular carcinoma. Sci China Life Sci 2018;61:727-8.
- Sack LM, Davoli T, Li MZ, Li Y, Xu Q, Naxerova K, *et al.* Profound tissue specificity in proliferation control underlies cancer drivers and aneuploidy patterns. Cell 2018;173:499-5.14E+25.
- 61. Goldstein SL. Nephrotoxicities. F1000Res 2017;6:55
- Kim EJ, Chen Y, Huang JQ, Li KM, Razmovski-Naumovski V, Poon J, et al. Evidence-based toxicity evaluation and scheduling of Chinese herbal medicines. J Ethnopharmacol 2013;146:40-61.
- Mehta RL, Awdishu L, Davenport A, Murray PT, Macedo E, Cerda J, et al. Phenotype standardization for drug-induced kidney disease. Kidney Int 2015;88:226-34.
- Kidney Disease: Improving Global Outcomes (KDIGO) Acute Kidney Injury Work Group. KDIGO clinical practice guideline for acute kidney injury. Kidney Int Suppl 2012;2:1-138.
- 65. Bellomo R, Ronco C, Kellum JA, Mehta RL, Palevsky P; Acute Dialysis Quality Initiative workgroup. Acute renal failure – Definition, outcome measures, animal models, fluid therapy and information technology needs: The Second International Consensus Conference of the Acute Dialysis Quality Initiative (ADQI) Group. Crit Care 2004;8:R204-12.
- Luyckx VA, Naicker S. Acute kidney injury associated with the use of traditional medicines. Nat Clin Pract Nephrol 2008;4:664-71.
- Allard T, Wenner T, Greten HJ, Efferth T. Mechanisms of herb-induced nephrotoxicity. Curr Med Chem 2013;20:2812-9.
- 68. Pickens CL, Milliron AR, Fussner AL, Dversdall BC, Langenstroer P, Ferguson S, *et al.* Abuse of guaifenesin-containing medications generates an excess of a carboxylate salt of beta-(2-methoxyphenoxy)-lactic acid, a guaifenesin metabolite, and results in urolithiasis. Urology 1999;54:23-7.
- Lebeau C, Debelle FD, Arlt VM, Pozdzik A, De Prez EG, Phillips DH, et al. Early proximal tubule injury in experimental aristolochic acid nephropathy: Functional and histological studies. Nephrol Dial Transplant 2005;20:2321-32.
- Yu Y, Zheng FL, Li H. Chinese herbs-induced renal failure with Fanconi syndrome: A report of 6 cases. Zhonghua Nei Ke Za Zhi 2003;42:110-2.
- Lee S, Lee T, Lee B, Choi H, Yang M, Ihm CG, *et al.* Fanconi syndrome and subsequent progressive renal failure caused by a Chinese herb containing aristolochic acid. Nephrology (Carlton) 2004;9:126-9.
- Hong YT, Fu LS, Chung LH, Hung SC, Huang YT, Chi CS, *et al.* Fanconi syndrome, interstitial fibrosis and renal failure by aristolochic acid in Chinese herbs. Pediatr Nephrol 2006;21:577-9.
- Pahwa R, Chatterjee VC. The toxicity of yellow oleander (Thevetia neriifolia juss) seed kernels to rats. Vet Hum Toxicol 1990;32:561-4.
- Samal K, Sahu H, Gopalakrishnakone P. Clinico-pathological study of *Thevetia peruviana* (yellow oleander) poisoning. J Wilderness Med 1992;3:382-6.
- Bandara V, Weinstein SA, White J, Eddleston M. A review of the natural history, toxinology, diagnosis and clinical management of *Nerium oleander* (common oleander) and *Thevetia peruviana* (yellow oleander) poisoning. Toxicon 2010;56:273-81.
- Samal KK, Sahu HK, Kar MK, Palit SK, Kar BC, Sahu CS, et al. Yellow oleander (*Cerbera thevetia*) poisoning with jaundice and renal failure. J Assoc Physicians India 1989;37:232-3.
- 77. Conz PA, La Greca G, Benedetti P, Bevilacqua PA, Cima L. Fucus vesiculosus: A nephrotoxic alga? Nephrol Dial Transplant 1998;13:526-7.
- Omar HR, Komarova I, El-Ghonemi M, Fathy A, Rashad R, Abdelmalak HD, *et al.* Licorice abuse: Time to send a warning message. Ther Adv Endocrinol Metab 2012;3:125-38.
- Berman JA, Setty A, Steiner MJ, Kaufman KR, Skotzko C. Complicated hypertension related to the abuse of ephedrine and caffeine alkaloids. J Addict Dis 2006;25:45-8.
- Polenakovic M, Stefanovic V. Balkan nephropathy. In: Davison A, Cameron J, Grunfeld J, editors. Oxford Textbook of Clinical Nephrology. New York: Oxford University Press; 1998. p. 1203.

- 81. Ivić M. Etiology of endemic nephropathy. Lijec Vjesn 1969;91:1273-81.
- Arlt VM, Ferluga D, Stiborova M, Pfohl-Leszkowicz A, Vukelic M, Ceovic S, *et al.* Is aristolochic acid a risk factor for Balkan endemic nephropathy-associated urothelial cancer? Int J Cancer 2002;101:500-2.
- Arlt VM, Stiborová M, vom Brocke J, Simões ML, Lord GM, Nortier JL, *et al.* Aristolochic acid mutagenesis: Molecular clues to the aetiology of Balkan endemic nephropathy-associated urothelial cancer. Carcinogenesis 2007;28:2253-61.
- 84. Schmeiser HH, Kucab JE, Arlt VM, Phillips DH, Hollstein M, Gluhovschi G, *et al.* Evidence of exposure to aristolochic acid in patients with urothelial cancer from a Balkan endemic nephropathy region of Romania. Environ Mol Mutagen 2012;53:636-41.
- Jelaković B, Karanović S, Vuković-Lela I, Miller F, Edwards KL, Nikolić J, *et al.* Aristolactam-DNA adducts are a biomarker of environmental exposure to aristolochic acid. Kidney Int 2012;81:559-67.
- Wojcikowski K, Wohlmuth H, Johnson DW, Gobe G. *Dioscorea* villosa (wild yam) induces chronic kidney injury via pro-fibrotic pathways. Food Chem Toxicol 2008;46:3122-31.
- Prud'homme GJ. Pathobiology of transforming growth factor beta in cancer, fibrosis and immunologic disease, and therapeutic considerations. Lab Invest 2007;87:1077-91.
- Liu YW, Shang HF, Wang CK, Hsu FL, Hou WC. Immunomodulatory activity of dioscorin, the storage protein of yam (*Dioscorea alata* cv. Tainong no 1) tuber. Food Chem Toxicol 2007;45:2312-8.
- 89. Wong Y, Qu S, Kong Q, Zhang X, Liang X, Hu Q, et al. Knowledge-based discovery of anti-fibrotic and pro-fibrotic activities from Chinese materia medica. In: Kuang X, editor. Recent Advances in Theories and Practice of Chinese Medicine. Rijeka, Croatia: Intech; 2012. p. 337-52.
- Sun R, Wu X, Liu, J, Sun L, Lv L. Experimental study of the rat renal toxicity of *Tripterygium wilfordii*, *Caulis aristolochiae* and *Leonurus*. Pharmacol Clin Chin Mater Med 2005;21:26-8.
- Sun R, Sun L, Wu X, Lv L. The attenuation effect of compound compatibility to nephrotoxicity of rat caused by Leonurus. Journal of Pharmacovigilance 2005;2:144-7.
- Lord GM, Cook T, Arlt VM, Schmeiser HH, Williams G, Pusey CD, et al. Urothelial malignant disease and Chinese herbal nephropathy. Lancet 2001;358:1515-6.
- Lord GM, Hollstein M, Arlt VM, Roufosse C, Pusey CD, Cook T, et al. DNA adducts and p53 mutations in a patient with aristolochic acid-associated nephropathy. Am J Kidney Dis 2004;43:e11-7.
- Cosyns JP, Jadoul M, Squifflet JP, Van Cangh PJ, van Ypersele de Strihou C. Urothelial malignancy in nephropathy due to Chinese herbs. Lancet 1994;344:188.
- Vanherweghem JL, Tielemans C, Simon J, Depierreux M. Chinese herbs nephropathy and renal pelvic carcinoma. Nephrol Dial Transplant 1995;10:270-3.
- Cosyns JP, Jadoul M, Squifflet JP, Wese FX, van Ypersele de Strihou C. Urothelial lesions in Chinese-herb nephropathy. Am J Kidney Dis 1999;33:1011-7.
- Nortier JL, Martinez MC, Schmeiser HH, Arlt VM, Bieler CA, Petein M, *et al.* Urothelial carcinoma associated with the use of a Chinese herb (*Aristolochia fangchi*) N Engl J Med 2000;342:1686-92.
- 98. Lemy A, Wissing KM, Rorive S, Zlotta A, Roumeguere T, Muniz Martinez MC, et al. Late onset of bladder urothelial carcinoma after kidney transplantation for end-stage aristolochic acid nephropathy: A case series with 15-year follow-up. Am J Kidney Dis 2008;51:471-7.
- Zlotta AR, Roumeguere T, Kuk C, Alkhateeb S, Rorive S, Lemy A, et al. Select screening in a specific high-risk population of patients suggests a stage migration toward detection of non-muscle-invasive bladder cancer. Eur Urol 2011;59:1026-31.
- 100. Chen CY, Liao YM, Tsai WM, Kuo HC. Upper urinary tract urothelial carcinoma in Eastern Taiwan: High proportion among all urothelial carcinomas and correlation with chronic kidney disease. J Formos Med Assoc 2007;106:992-8.
- 101. Chen CH, Dickman KG, Moriya M, Zavadil J, Sidorenko VS, Edwards KL, *et al.* Aristolochic acid-associated urothelial cancer in Taiwan. Proc Natl Acad Sci U S A 2012;109:8241-6.
- 102. Hoang ML, Chen CH, Sidorenko VS, He J, Dickman KG, Yun BH, et al. Mutational signature of aristolochic acid exposure as revealed by whole-exome sequencing. Sci Transl Med 2013;5:197ra102.

- 103. Poon SL, Pang ST, McPherson JR, Yu W, Huang KK, Guan P, *et al.* Genome-wide mutational signatures of aristolochic acid and its application as a screening tool. Sci Transl Med 2013;5:197ra101.
- 104. Scelo G, Riazalhosseini Y, Greger L, Letourneau L, Gonzàlez-Porta M, Wozniak MB, *et al.* Variation in genomic landscape of clear cell renal cell carcinoma across Europe. Nat Commun 2014;5:5135.
- 105. Turesky RJ, Yun BH, Brennan P, Mates D, Jinga V, Harnden P, et al. Aristolochic acid exposure in Romania and implications for renal cell carcinoma. Br J Cancer 2016;114:76-80.
- 106. Yun BH, Rosenquist TA, Sidorenko V, Iden CR, Chen CH, Pu YS, et al. Biomonitoring of aristolactam-DNA adducts in human tissues using ultra-performance liquid chromatography/ion-trap mass spectrometry. Chem Res Toxicol 2012;25:1119-31.
- 107. Jelaković B, Castells X, Tomić K, Ardin M, Karanović S, Zavadil J, et al. Renal cell carcinomas of chronic kidney disease patients harbor the mutational signature of carcinogenic aristolochic acid. Int J Cancer 2015;136:2967-72.
- Huxtable RJ. The harmful potential of herbal and other plant products. Drug Saf 1990;5 Suppl 1:126-36.
- Edwards IR, Aronson JK. Adverse drug reactions: Definitions, diagnosis, and management. Lancet 2000;356:1255-9.
- 110. Kim LA, Amarnani D, Gnanaguru G, Tseng WA, Vavvas DG, D'Amore PA, *et al.* Tamoxifen toxicity in cultured retinal pigment epithelial cells is mediated by concurrent regulated cell death mechanisms. Invest Ophthalmol Vis Sci 2014;55:4747-58.
- Wallach D, Kang TB, Dillon CP, Green DR. Programmed necrosis in inflammation: Toward identification of the effector molecules. Science 2016;352:aaf2154.
- 112.Sin XY. A reference on poisonous and adverse effects of Chinese medicine. Available from: http://www.xys.org/xys/netters/Fang-Zhouzi/ sohu/zhongyao.txt. [Last accessed on 2017 May 18].
- 113.Xing SM. A private library. Available from: http://www. 360doc.com/ content/16/0620/18/15889276_569330403.shtml. [Last accessed on 2017 May 18].
- 114. Zhang C, Ai C. Common nephrotoxic TCM herbs. Zhong Guo Min Jian Liao Fa 2006;14:35-6.
- 115. Rueda DC, Zaugg J, Quitschau M, Reich E, Hering S, Hamburger M, et al. Discovery of GABA (A) receptor modulator aristolactone in a commercial sample of the Chinese herbal drug "Chaihu" (*Bupleurum* chinense roots) unravels adulteration by nephrotoxic Aristolochia manshuriensis roots. Planta Med 2012;78:207-10.
- Wang XK, Zhao Y, Liu T, Yi Y, Li CY, Wang HJ, *et al.* Ninety-day subchronic oral toxicity study of *Senecio scandens* extract in rats. Biol Pharm Bull 2015;38:1548-56.
- 117. Wang D, Huang L, Chen S. *Senecio scandens* Buch.-Ham.: A review on its ethnopharmacology, phytochemistry, pharmacology, and toxicity. J Ethnopharmacol 2013;149:1-23.
- 118. Naranjo CA, Busto U, Sellers EM, Sandor P, Ruiz I, Roberts EA, et al. A method for estimating the probability of adverse drug reactions. Clin Pharmacol Ther 1981;30:239-45.
- Davies E, Rowe P, James S, Pirmohamed M. An investigation of disagreement in causality assessment of adverse drug reactions. Pharm Med 2011;25:17-24.
- Rosemarin A, Notini M, Holmgren K. The fate of arsenic in the (c) Baltic Sea. *Fucus vesiculosus* ecosystem. Ambio 1985;6:342-5.
- Walkiw O, Douglas DE. Health food supplements prepared from kelp-A source of elevated urinary arsenic. Clin Toxicol 1975;8:325-31.
- 122. Wu MS, Hong JJ, Lin JL, Yang CW, Chien HC. Multiple tubular dysfunction induced by mixed Chinese herbal medicines containing

cadmium. Nephrol Dial Transplant 1996;11:867-70.

- 123. Chan K, Shaw D, Simmonds MS, Leon CJ, Xu Q, Lu A, *et al.* Good practice in reviewing and publishing studies on herbal medicine, with special emphasis on traditional Chinese medicine and Chinese materia medica. J Ethnopharmacol 2012;140:469-75.
- 124. Rivera D, Allkin R, Obón C, Alcaraz F, Verpoorte R, Heinrich M. What is in a name? The need for accurate scientific nomenclature for plants. J Ethnopharmacol. 2014;152:393-402.
- 125. Royal Botanic Gardens Kew. Medicinal Plant Names Services. Available from: https://www.kew.org/science/data-and-resources/ tools-and-services/medicinal-plant-names-services. [Last accessed on 2019 Feb 20].
- 126. The Uppsala Monitoring Centre. The Herbal Anatomical Therapeutic Chemical Classification System. Available from: https://www.whoumc.org/whodrug/whodrug-portfolio/whodrug-global/herbal-atc/. [Last accessed on 2019 Feb 20].
- 127. Endre ZH, Kellum JA, Di Somma S, Doi K, Goldstein SL, Koyner JL, et al. Differential diagnosis of AKI in clinical practice by functional and damage biomarkers: Workgroup statements from the tenth acute dialysis quality initiative consensus conference. Contrib Nephrol 2013;182:30-44.
- Endre ZH, Pickering JW, Walker RJ. Clearance and beyond: The complementary roles of GFR measurement and injury biomarkers in acute kidney injury (AKI). Am J Physiol Renal Physiol 2011;301:F697-707.
- 129. Ronco C, Legrand M, Goldstein SL, Hur M, Tran N, Howell EC, et al. Neutrophil gelatinase-associated lipocalin: Ready for routine clinical use? An international perspective. Blood Purif 2014;37:271-85.
- Otterbein LE, Choi AM. Heme oxygenase: Colors of defense against cellular stress. Am J Physiol Lung Cell Mol Physiol 2000;279:L1029-37.
- 131. Poss KD, Tonegawa S. Heme oxygenase 1 is required for mammalian iron reutilization. Proc Natl Acad Sci U S A 1997;94:10919-24.
- 132. Poss KD, Tonegawa S. Reduced stress defense in heme oxygenase 1-deficient cells. Proc Natl Acad Sci U S A 1997;94:10925-30.
- 133. Agarwal A, Balla J, Alam J, Croatt AJ, Nath KA. Induction of heme oxygenase in toxic renal injury: A protective role in cisplatin nephrotoxicity in the rat. Kidney Int 1995;48:1298-307.
- 134. Nath KA, Balla G, Vercellotti GM, Balla J, Jacob HS, Levitt MD, *et al.* Induction of heme oxygenase is a rapid, protective response in rhabdomyolysis in the rat. J Clin Invest 1992;90:267-70.
- 135. Vogt BA, Shanley TP, Croatt A, Alam J, Johnson KJ, Nath KA, et al. Glomerular inflammation induces resistance to tubular injury in the rat. A novel form of acquired, heme oxygenase-dependent resistance to renal injury. J Clin Invest 1996;98:2139-45.
- Zager R, Johnson A, Becker K. Plasma and urinary heme oxygenase-1 in AKI. J Am Soc Nephrol 2012;23:1048-57.
- 137. Adler M, Ramm S, Hafner M, Muhlich JL, Gottwald EM, Weber E, et al. A quantitative approach to screen for nephrotoxic compounds in vitro. J Am Soc Nephrol 2016;27:1015-28.
- 138. Xu Q, Feng Y, Duez P, Hendry B, Hylands P. The hunt for anti-fibrotic and pro-fibrotic botanicals. Science 2014;346 Suppl 6216:S19-20.
- 139. Pokladnikova J, Meyboom RH, Meincke R, Niedrig D, Russmann S. Allergy-like immediate reactions with herbal medicines: A retrospective study using data from VigiBase®. Drug Saf 2016;39:455-64.
- Mullard A. FDA unveils searchable adverse events system. Nat Rev Drug Discov 2017;16:743.

Supplementary Table 1: Botanicals associated with the acute kidney injury phenotype of botanical-induced kidney injury

Botanical	Nephrotoxic manifestations	References and levels of evidence
Aristolochia spp.	AKI (ATN or AIN), CKD and	Epidemiological survey ^[1]
	tubular disorder	Animal model ^[2]
		Case-control ^[3]
		Cohort study ^[4]
Callilepis laureola and other atractyloside-containing	AKI (ATN)	Case report ^[5]
spp.		Case report ^[6]
		Retrospective study ^[7]
Cupressus funebris, Taxus celebica, and other	AKI (AIN or ATN)	Case report ^[8]
nephrotoxic flavonoids-containing spp.		Case report ^[9]
Tripterygium wilfordii Hook F	AKI (ATN)	Animal model ^[10]
		Animal model ^[11]
		Case report ^[12]
		Animal model ^[13]
		Animal model ^[14]
Aloe spp.	AKI (AIN)	Case report ^[15]
		Animal model ^[16]
Teucrium polium	AKI (ATN)	Animal model ^[17]
Artemisia absinthium (wormwood)	AKI	Case report ^[18]
Uncaria tomentosa or Uncaria guianensis (cat's claw)	AKI (AIN)	Case report ^[19]

AKI: Acute kidney injury, ATN: Acute tubular necrosis, AIN: Acute interstitial nephritis, CKD: Chronic kidney disease

Supplementary Table 2: Botanicals associated with the nephrolithiasis phenotype of botanical-induced kidney injury

Botanical	Nephrotoxic manifestations	Reference and levels of evidence
Vaccinium spp. (Cranberry)	Nephrolithiasis	RCT ^[20]
		Cohort study ^[21]
Rheum officinale (Rhubarb)	Nephrolithiasis (may lead to AKI)	Case report ^[22]
Averrhoa carambola (Star fruit)	Nephrolithiasis (may lead to AKI)	Case report ^[23]
		Animal model ^[24]
		Animal model ^[25]
		Case report ^[26]
		<i>In vitro</i> evidence ^[27]
		<i>In vitro</i> evidence ^[28]
Herniaria hirsuta L. (Hairy	Nephrolithiasis	Animal model ^[29]
Rupture Wort)		<i>In vitro</i> evidence ^[30]
		Animal model ^[31]
Ephedra sinica	Nephrolithiasis	Case report ^[32]
Guaiacum officinale		Systematic review ^[33]
		Case report ^[34]
		Case report ^[35]
		Case report ^[36]
		Cohort study ^[37]
		Case report ^[38]

RCT: Randomized controlled trial, AKI: Acute kidney injury

Supplementary Table 3: Botanicals associated with the tubular dysfunction phenotype of botanical-induced kidney injury

Botanical	Nephrotoxic manifestations	References and levels of evidence
Aristolochia spp.	Tubular dysfunction, which can manifest as Fanconi	Animal model ^[39]
	syndrome	Cohort study ^[40]
		Case report ^[41]
		Case report ^[42]
Juniperus communis (Juniper berries)	Tubular dysfunction, which can manifest as diuresis, etc.	Animal model ^[43]
Fucus vesiculosus (Bladderwrack)	Tubular dysfunction, characterized by degeneration	Case report ^[44]
Cleistanthus collinus	Tubular dysfunction, characterized by distal renal	Cohort study ^[45]
tubular acid	tubular acidosis	Animal model ^[46]
		Case report ^[47]
		Case report ^[48]

Mechanism of nephrotoxicity	Plant species	Nephrotoxic manifestation and pathogenesis	Reference and level of evidence
Inducing cell	Aristolochia spp.	AKI: Inducing apoptosis and interfering with cell cycle in	In vitro ^[49]
death		tubular cells	In vitro ^[50]
			Animal model ^[51]
			In vitro ^[52]
	Averrhoa carambola	AKI: Focal tubular deposition of crystals inducing apoptosis	In vitro ^[24]
	(star fruit)	with fragmentation and cytoplasmic vacuolization	In vitro ^[25]
			In vitro ^[27]
			In vitro ^[28]
	Atractyloside-containing	AKI: Inhibition of mitochondrial ATP synthesis; mitochondrial	Animal model ^[53]
	spp.	membrane permeability pore activation	In vitro ^[54]
			Animal model ^[55]
			Animal model ^[56]
	Tripterygium wilfordii	AKI: Inducing oxidative stress, reducing superoxide dismutase and glutathione peroxidase	Animal model ^[14]
	Cleistanthus collinus	AKI and tubular dysfunction: Inhibition of cell division and	In vitro ^[57]
		DNA synthesis; reduction of glutathione and ATPases, leading to	In vitro ^[58]
		oxidative stress	In vitro ^[59]
			In vitro ^[60]
			In vitro ^[61]
	Aconitum spp., e.g., Aconitum carmichaelii or Allium tanguticum	AKI: Inhibiting the tricarboxylic acid cycle in myocardium, leading to renal ischemia and hypoxia, releasing oxidative stressors which activate proapoptotic genes in renal cells	Animal model ^[62]
Blockade of renal transport	Harpagophytum procumbens (Devil's claw)	AKI: Downregulates P-glycoprotein transporter expression and activity	In vitro ^[63]
processes	Tripterygium wilfordii Hook	Tubular dysfunction: Inhibits specific segments of organic anion transporters, required for secretion and absorption	Animal model ^[64]
Crystal and stone	Vaccinium spp. (cranberry)	Nephrolithiasis: Contains high concentrations of oxalate	RCT ^[20]
formation			Cohort study ^[21]
	Averrhoa carambola (star	Nephrolithiasis and AKI: Focal tubular deposition of calcium	Case report ^[22]
	fruit) and Rheum officinale	oxalate crystals due to high oxalic acid content	Case report ^[23]
	(rhubarb)		Animal model ^[24]
			Animal model ^[25]
			Case report ^[26]
			In vitro ^[27]
			In vitro ^[28]
			Literature review[65]

Supplementary Table 4: A summary of proposed mechanisms of botanical-induced kidney injury

Supplementary Table 4: Contd				
Mechanism of nephrotoxicity	Plant species	Nephrotoxic manifestation and pathogenesis	Reference and level of evidence	
	Ephedra sinica; Guaiacum officinale	Nephrolithiasis: Poor solubility leads to precipitation out of the urine when consumed in large quantities	Case report ^[32] Systematic review ^[33] Case report ^[34] Case report ^[35] Case report ^[36] Cohort study ^[37] Case report ^[38] Case report ^[66]	
Carcinogenesis	Aristolochia spp.	UUC/RCC: AA-derived ions form covalent adducts with DNA purine bases, causing TP53 mutation	Molecular and epidemiological evidence ^[67-71]	
Hypertension	Glycyrrhiza glabra	CKD: Inhibition of renal 11B-hydroxysteroid dehydrogenase, leading to inappropriate activation of mineralocorticoid receptors	Case report ^[72] Animal model ^[73] <i>In vitro</i> ^[74] Clinical study ^[75] Animal model ^[76] Animal model ^[77]	
	<i>Ephedra</i> spp.	CKD: Sympathomimetic activity	Case report ^[78] RCT ^[79]	
Idiosyncratic	Uncaria tomentosa	AKI: Type B IgE-mediated allergic reaction	Case report ^[19]	

AKI: Acute kidney injury, UUC: Upper urinary tract urothelial cancer, RCC: Renal cell carcinoma, CKD: Chronic kidney disease, IgE: Immunoglobulin E, AA: Aristolochic acid, RCT: Randomized controlled trial, ATP: Adenosine triphosphate

REFERENCES TO **S**UPPLEMENTARY **T**ABLES

- 1. Vanherweghem JL, Depierreux M, Tielemans C, Abramowicz D, Dratwa M, Jadoul M, *et al.* Rapidly progressive interstitial renal fibrosis in young women: Association with slimming regimen including Chinese herbs. Lancet 1993;341:387-91.
- Schmeiser HH, Bieler CA, Wiessler M, van Ypersele de Strihou C, Cosyns JP. Detection of DNA adducts formed by aristolochic acid in renal tissue from patients with Chinese herbs nephropathy. Cancer Res 1996;56:2025-8.
- Nortier JL, Martinez MC, Schmeiser HH, Arlt VM, Bieler CA, Petein M, et al. Urothelial carcinoma associated with the use of a Chinese herb (*Aristolochia fangchi*) N Engl J Med 2000;342:1686-92.
- Bieler CA, Stiborova M, Wiessler M, Cosyns JP, van Ypersele de Strihou C, Schmeiser HH. 32P-post-labelling analysis of DNA adducts formed by aristolochic acid in tissues from patients with Chinese herbs nephropathy. Carcinogenesis 1997;18:1063-7.
- Steenkamp V, Stewart MJ, Zuckerman M. Detection of poisoning by impila (*Callilepis laureola*) in a mother and child. Hum Exp Toxicol 1999;18:594-7.
- Seedat YK, Hitchcock PJ. Acute renal failure from *Callilepsis laureola*. S Afr Med J 1971;45:832-3.
- Watson AR, Coovadia HM, Bhoola KD. The clinical syndrome of impila (*Callilepis laureola*) poisoning in children. S Afr Med J 1979;55:290-2.
- Lee JJ, Chen HC. Flavonoid-induced acute nephropathy by *Cupressus funebris* endl (Mourning cypress). Am J Kidney Dis 2006;48:e81-5.
- Lin JL, Ho YS. Flavonoid-induced acute nephropathy. Am J Kidney Dis 1994;23:433-40.
- Shamon LA, Pezzuto JM, Graves JM, Mehta RR, Wangcharoentrakul S, Sangsuwan R, *et al.* Evaluation of the mutagenic, cytotoxic, and antitumor potential of triptolide, a highly oxygenated diterpene isolated from *Tripterygium wilfordii*. Cancer Lett 1997;112:113-7.
- Liu L, Wang Z, Huang G, Liu Y. The influence of triptolide sub-chronic intoxication on kidney and testicle in mice. J Huazhong Univ Sci Technol Med Sci 2001;30:214-7.
- Chou WC, Wu CC, Yang PC, Lee YT. Hypovolemic shock and mortality after ingestion of *Tripterygium wilfordii* hook F.: A case report. Int J Cardiol 1995;49:173-7.
- 13. Li XX, Du FY, Liu HX, Ji JB, Xing J. Investigation of the active

components in *Tripterygium wilfordii* leading to its acute hepatotoxicty and nephrotoxicity. J Ethnopharmacol 2015;162:238-43.

- Yang F, Ren L, Zhuo L, Ananda S, Liu L. Involvement of oxidative stress in the mechanism of triptolide-induced acute nephrotoxicity in rats. Exp Toxicol Pathol 2012;64:905-11.
- Luyckx VA, Ballantine R, Claeys M, Cuyckens F, Van den Heuvel H, Cimanga RK, *et al.* Herbal remedy-associated acute renal failure secondary to cape aloes. Am J Kidney Dis 2002;39:E13.
- Koroye O, Siminialayi I, Etebu E. Effects of oral administration of aloe vera plus on the heart and kidney: A subacute toxicity study in rat models. Niger Health J 2010;10:1-2.
- Baradaran A, Madihi Y, Merrikhi A, Rafieian-Kopaei M, Nematbakhsh M, Asgari A, *et al.* Nephrotoxicity of hydroalcoholic extract of *Teucrium polium* in Wistar rats. Pak J Med Sci 2013;29:329-33.
- Weisbord SD, Soule JB, Kimmel PL. Poison on line Acute renal failure caused by oil of wormwood purchased through the internet. N Engl J Med 1997;337:825-7.
- Hilepo JN, Bellucci AG, Mossey RT. Acute renal failure caused by 'cat's claw' herbal remedy in a patient with systemic lupus erythematosus. Nephron 1997;77:361.
- Gettman MT, Ogan K, Brinkley LJ, Adams-Huet B, Pak CY, Pearle MS, et al. Effect of cranberry juice consumption on urinary stone risk factors. J Urol 2005;174:590-4.
- Terris MK, Issa MM, Tacker JR. Dietary supplementation with cranberry concentrate tablets may increase the risk of nephrolithiasis. Urology 2001;57:26-9.
- 22. Albersmeyer M, Hilge R, Schröttle A, Weiss M, Sitter T, Vielhauer V. Acute kidney injury after ingestion of rhubarb: Secondary oxalate nephropathy in a patient with type 1 diabetes. BMC Nephrol 2012;13:141.
- Chen CL, Fang HC, Chou KJ, Wang JS, Chung HM. Acute oxalate nephropathy after ingestion of star fruit. Am J Kidney Dis 2001;37:418-22.
- Fang HC, Lee PT, Lu PJ, Chen CL, Chang TY, Hsu CY, et al. Mechanisms of star fruit-induced acute renal failure. Food Chem Toxicol 2008;46:1744-52.
- Fang HC, Chen CL, Wang JS, Chou KJ, Chiou YS, Lee PT, *et al.* Acute oxalate nephropathy induced by star fruit in rats. Am J Kidney Dis 2001;38:876-80.

- Niticharoenpong K, Chalermsanyakorn P, Panvichian R, Kitiyakara C. Acute deterioration of renal function induced by star fruit ingestion in a patient with chronic kidney disease. J Nephrol 2006;19:682-6.
- Koul S, Fu S, Koul H. Oxalate exposure promotes reinitiation of the DNA synthesis and apoptosis of HK-2 cells, a line of human renal epithelial cells. Ann N Y Acad Sci 2003;1010:292-5.
- Miller C, Kennington L, Cooney R, Kohjimoto Y, Cao LC, Honeyman T, et al. Oxalate toxicity in renal epithelial cells: Characteristics of apoptosis and necrosis. Toxicol Appl Pharmacol 2000;162:132-41.
- Atmani F, Slimani Y, Mimouni M, Hacht B. Prophylaxis of calcium oxalate stones by *Herniaria hirsuta* on experimentally induced nephrolithiasis in rats. BJU Int 2003;92:137-40.
- Atmani F, Khan SR. Effects of an extract from *Herniaria hirsuta* on calcium oxalate crystallization *in vitro*. BJU Int 2000;85:621-5.
- Atmani F, Slimani Y, Mimouni M, Aziz M, Hacht B, Ziyyat A, et al. Effect of aqueous extract from *Herniaria hirsuta* L. On experimentally nephrolithiasic rats. J Ethnopharmacol 2004;95:87-93.
- Powell T, Hsu FF, Turk J, Hruska K. Ma-huang strikes again: Ephedrine nephrolithiasis. Am J Kidney Dis 1998;32:153-9.
- Bennett S, Hoffman N, Monga M. Ephedrine- and guaifenesin-induced nephrolithiasis. J Altern Complement Med 2004;10:967-9.
- Blau J. Ephedrine nephrolithiasis associated with chronic ephedrine abuse. J Urol 1998;160:825.
- Small E, Sandefur BJ. Acute renal failure after ingestion of guaifenesin and dextromethorphan. J Emerg Med 2014;47:26-9.
- Whelan C, Schwartz BF. Bilateral guaifenesin ureteral calculi. Urology 2004;63:175-6.
- Assimos DG, Langenstroer P, Leinbach RF, Mandel NS, Stern JM, Holmes RP. Guaifenesin- and ephedrine-induced stones. J Endourol 1999;13:665-7.
- Smith CL, Gemar SK, Lewis MJ. Pseudoephedrine urolithiasis associated with acute renal failure. Nephrol Dial Transplant 2004;19:263-4.
- Lebeau C, Debelle FD, Arlt VM, Pozdzik A, De Prez EG, Phillips DH, et al. Early proximal tubule injury in experimental aristolochic acid nephropathy: Functional and histological studies. Nephrol Dial Transplant 2005;20:2321-32.
- Yu Y, Zheng FL, Li H. Chinese herbs-induced renal failure with Fanconi syndrome: A report of 6 cases. Zhonghua Nei Ke Za Zhi 2003;42:110-2.
- 41. Lee S, Lee T, Lee B, Choi H, Yang M, Ihm CG, *et al.* Fanconi syndrome and subsequent progressive renal failure caused by a Chinese herb containing aristolochic acid. Nephrology (Carlton) 2004;9:126-9.
- Hong YT, Fu LS, Chung LH, Hung SC, Huang YT, Chi CS, et al. Fanconi syndrome, interstitial fibrosis and renal failure by aristolochic acid in Chinese herbs. Pediatr Nephrol 2006;21:577-9.
- Stanic G, Samarzija I, Blazevic N. Time-dependent diuretic response in rats treated with Juniper berry preparations. Phytothe Res 1998;12:494-7.
- 44. Conz PA, La Greca G, Benedetti P, Bevilacqua PA, Cima L. Fucus vesiculosus: A nephrotoxic alga? Nephrol Dial Transplant 1998;13:526-7.
- Nampoothiri K, Chrispal A, Begum A, Jasmine S, Gopinath KG, Zachariah A. A clinical study of renal tubular dysfunction in *Cleistanthus collinus* (Oduvanthalai) poisoning. Clin Toxicol (Phila) 2010;48:193-7.
- 46. Maneksh D, Sidharthan A, Kettimuthu K, Kanthakumar P, Lourthuraj AA, Ramachandran A, *et al.* Cleistanthus collinus induces type I distal renal tubular acidosis and type II respiratory failure in rats. Indian J Pharmacol 2010;42:178-84.
- Benjamin SP, Fernando ME, Jayanth JJ, Preetha B. Cleistanthus collinus poisoning. J Assoc Physicians India 2006;54:742-4.
- Das S, Hamide A, Mohanty MK, Muthusamy R. Fatal *Cleistanthus collinus* toxicity: A case report and review of literature. J Forensic Sci 2014;59:1441-7.
- Romanov V, Whyard TC, Waltzer WC, Grollman AP, Rosenquist T. Aristolochic acid-induced apoptosis and G2 cell cycle arrest depends on ROS generation and MAP kinases activation. Arch Toxicol 2015;89:47-56.
- Yu FY, Wu TS, Chen TW, Liu BH. Aristolochic acid I induced oxidative DNA damage associated with glutathione depletion and ERK1/2 activation in human cells. Toxicol *In Vitro* 2011;25:810-6.
- Bonventre JV. Primary proximal tubule injury leads to epithelial cell cycle arrest, fibrosis, vascular rarefaction, and glomerulosclerosis.

Kidney Int Suppl (2011) 2014;4:39-44.

- 52. Gao R, Zheng F, Liu Y, Zheng D, Li X, Bo Y, et al. Aristolochic acid I-induced apoptosis in LLC-PK1 cells and amelioration of the apoptotic damage by calcium antagonist. Chin Med J (Engl) 2000;113:418-24.
- Obatomi DK, Bach PH. Inhibition of mitochondrial respiration and oxygen uptake in isolated rat renal tubular fragments by atractyloside. Toxicol Lett 1996;89:155-61.
- Vancompernolle K, Van Herreweghe F, Pynaert G, Van de Craen M, De Vos K, Totty N, *et al.* Atractyloside-induced release of cathepsin B, a protease with caspase-processing activity. FEBS Lett 1998;438:150-8.
- 55. Carpenedo F, Luciani S, Scaravilli F, Palatini P, Santi R. Nephrotoxic effect of atractyloside in rats. Arch Toxicol 1974;32:169-80.
- Koechel DA, Krejci ME. Extrarenal and direct renal actions of atractyloside contribute to its acute nephrotoxicity in pentobarbital-anesthetized dogs. Toxicology 1993;79:45-66.
- Pradheepkumar CP, Panneerselvam N, Shanmugam G. Cleistanthin A causes DNA strand breaks and induces apoptosis in cultured cells. Mutat Res 2000;464:185-93.
- Kumar CP, Pande G, Shanmugam G. Cleistanthin B causes G1 arrest and induces apoptosis in mammalian cells. Apoptosis 1998;3:413-9.
- Prabhakaran C, Kumar P, Panneerselvam N, Rajesh S, Shanmugam G. Cytotoxic and genotoxic effects of cleistanthin B in normal and tumour cells. Mutagenesis 1996;11:553-7.
- Kettimuthu K, Ramachandran A, Lourthuraj A, Manickam S, Subramani S. Mechanism of toxicity of *Cleistanthus collinus*: Vacuolar H+ATPases are a putative target. Clin Toxicol 2009;47:724.
- Sørensen MG, Henriksen K, Neutzsky-Wulff AV, Dziegiel MH, Karsdal MA. Diphyllin, a novel and naturally potent V-ATPase inhibitor, abrogates acidification of the osteoclastic resorption lacunae and bone resorption. J Bone Miner Res 2007;22:1640-8.
- Lei H, Yi J. Observation of apoptosis in renal tubule epithelial cell after aconitine poisoning. Ind Health Occup Dis 2005;2:5.
- Romiti N, Tramonti G, Corti A, Chieli E. Effects of devil's claw (*Harpagophytum procumbens*) on the multidrug transporter ABCB1/P-glycoprotein. Phytomedicine 2009;16:1095-100.
- Dan H, Peng RX, Ao Y, Liu YH. Segment-specific proximal tubule injury in *Tripterygium* glycosides intoxicated rats. J Biochem Mol Toxicol 2008;22:422-8.
- Massey LK, Roman-Smith H, Sutton RA. Effect of dietary oxalate and calcium on urinary oxalate and risk of formation of calcium oxalate kidney stones. J Am Diet Assoc 1993;93:901-6.
- Dang H, Raut R, Shih W, Lisawat P. Recurrent nephrolithiasis secondary to guaifenesin abuse. Am J Kidney Dis 2015;65:29. Available from: https://www.ajkd.org/article/S0272-6386(15)00151-1/pdf. [Last accessed online on 2019 Feb 02].
- Jelaković B, Karanović S, Vuković-Lela I, Miller F, Edwards KL, Nikolić J, *et al.* Aristolactam-DNA adducts are a biomarker of environmental exposure to aristolochic acid. Kidney Int 2012;81:559-67.
- Chen CH, Dickman KG, Moriya M, Zavadil J, Sidorenko VS, Edwards KL, *et al.* Aristolochic acid-associated urothelial cancer in Taiwan. Proc Natl Acad Sci U S A 2012;109:8241-6.
- Scelo G, Riazalhosseini Y, Greger L, Letourneau L, Gonzàlez-Porta M, Wozniak MB, *et al.* Variation in genomic landscape of clear cell renal cell carcinoma across Europe. Nat Commun 2014;5:5135.
- Turesky RJ, Yun BH, Brennan P, Mates D, Jinga V, Harnden P, *et al.* Aristolochic acid exposure in Romania and implications for renal cell carcinoma. Br J Cancer 2016;114:76-80.
- Yun BH, Rosenquist TA, Sidorenko V, Iden CR, Chen CH, Pu YS, *et al.* Biomonitoring of aristolactam-DNA adducts in human tissues using ultra-performance liquid chromatography/ion-trap mass spectrometry. Chem Res Toxicol 2012;25:1119-31.
- 72. Omar HR, Komarova I, El-Ghonemi M, Fathy A, Rashad R, Abdelmalak HD, *et al.* Licorice abuse: Time to send a warning message. Ther Adv Endocrinol Metab 2012;3:125-38.
- Whorwood CB, Sheppard MC, Stewart PM. Licorice inhibits 11 beta-hydroxysteroid dehydrogenase messenger ribonucleic acid levels and potentiates glucocorticoid hormone action. Endocrinology 1993;132:2287-92.
- 74. Calò LA, Zaghetto F, Pagnin E, Davis PA, De Mozzi P, Sartorato P, et al. Effect of aldosterone and glycyrrhetinic acid on the protein expression

of PAI-1 and p22(phox) in human mononuclear leukocytes. J Clin Endocrinol Metab 2004;89:1973-6.

- Stewart PM, Wallace AM, Valentino R, Burt D, Shackleton CH, Edwards CR, *et al.* Mineralocorticoid activity of liquorice: 11-beta-hydroxysteroid dehydrogenase deficiency comes of age. Lancet 1987;2:821-4.
- Kageyama Y, Suzuki H, Saruta T. Role of glucocorticoid in the development of glycyrrhizin-induced hypertension. Clin Exp Hypertens 1994;16:761-78.
- Gomez-Sanchez EP, Gomez-Sanchez CE. Central hypertensinogenic effects of glycyrrhizic acid and carbenoxolone. Am J Physiol 1992;263:E1125-30.
- Berman JA, Setty A, Steiner MJ, Kaufman KR, Skotzko C. Complicated hypertension related to the abuse of ephedrine and caffeine alkaloids. J Addict Dis 2006;25:45-8.
- Haller CA, Jacob P, Benowitz NL. Short-term metabolic and hemodynamic effects of ephedra and Guarana combinations. Clin Pharmacol Ther 2005;77:560-71.